Metodu ile Değerlendirme İmkanları. İ.Ü Orman Fakültesi Yayınları No. 2326/231, İstanbul (1978). — Tank, T.: Selüloz Üretimi Bakımından Doğu Çınarı Odununun Bazı Özellikleri Üzerine Araştırmalar. İ.Ü Orman Fakültesi Yayın No. 2669/290, İstanbul (1982). — Tunçtaner, K.: Research Works on Improvement of Poplar, Willow, and Fast Growing Exotic Coniferous Species. FO: TCP/TUR/6651, Training Document No. 6, İzmit (1987). — Tunçtaner, K.: Kavak Genetik Seleksiyon Çalışmalarında Gelişmeler ve İthal Edilen Bazı Yeni Kavak Klonlarının Fidanlık Performansları ile İlgili Karşılaştırmalar. Kavak ve Hızlı Gelişen Orman Ağaçları Araştırma Enstitüsü Dergisi, İzmit (1988). — Urgenç, S.: Orman Ağaçları İslahı. İ.Ü Orman Fakültesi Yayın No. 2836/293, İstanbul (1982). — Valle, G.: Mini-Rotations de Taillis de Peuplier, Sources de Fourrage, d'energie et de Re Venus Pour les Com-

munautes Rurales du Quebec. Pre'pare Pour la XVI. Session de la Commission Internationale du Peuplier, İzmir (1980). — VIART, M.: Mini-Monagraphie Sur Les Saules. Technical Consultation on Fast Growing Plantation Broadleaved Trees for Mediterranean and temperate Zones. Lisbon, Portugal (1979). — VIART, M.: Association of Poplar and Willow Cultivation with Agriculture for the Benefit of Rural Communities. FO: CIP/80/3 (1980). — WASIELEWSKI, D. H.: Cultivation of Willow in Central and South Eastern Europe. University of Agricultural Science. Energy Forest Project, Uppsala, Sweden (1982). — YALTIRIK, F.: Dendroloji Ders Kitabı II. Angiospermae (Kapalı Tohumlular). İ.Ü Orman Fakültesi Yayın No. 3509/390. İstanbul (1988). — ZSUFFA, L.: Vegetative Propagation of Cottonwood by Rooting Cuttings. Proceedings; Symposium on Eastern Cottonwood and Related Species. Greenville, MS. (1976).

Fertility Variation and Gene Diversity in Clonal Seed Orchards of Pinus brutia, Pinus nigra and Pinus sylvestris in Turkey

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Summary

Clonal variation in the production of female and male strobili was studied in seed orchards of Pinus brutia in southern, and those of P. nigra and P. sylvestris in northern Turkey. Large differences in female and male fertility among clones were found. The most abundant quartile of clones in P. brutia, P. nigra and P. sylvestris seed orchards produced 44.1%, 40.5%, 33.9% and 37.8% of total female, and $76.1\%,\,62.6\%,\,36.7\%$ and 34.1% in total male strobilus production, respectively. There were positive correlations between female and male strobilus production in all seed orchards. The status numbers (N_s) were calculated as 19.3 (64% of census number) and 23.2 (66%) in two P. brutia seed orchards, 29.2 (94%) in a P. nigra seed orchard and 25.7 (92%) in a P. sylvestris seed orchard. Male fertility variation was larger than female fertility variation in P. brutia and P. nigra seed orchards, while it was an opposite situation in the P. sylvestris seed orchard. The effect of fertility variation was discussed for the establishment of new seed orchards and the management of current orchards in Turkey.

Key words: Pinus brutia, Pinus nigra, Pinus sylvestris, seed orchard, fertility variation, status number, gene diversity.

Introduction

A seed orchard is an important seed source for forest plantation, and it keeps generally high genetic diversity compared to natural forests where orchard parents were selected (SAVOLINEN and Kärkkäinen, 1992). *Pinus brutia* Ten., *P. nigra* ARNOLD. and *P. sylvestris* L. are among most important native *Pinus* species in Turkey, and they occupy about 3.7 (26.4%), 2.5 (17.9%) and 0.7 (5.0%) million ha of the total 14 million ha of high forest area in Turkey, respectively (CALISKAN, 1998).

Large clonal differences in female and male fertility have been reported in seed orchards (Kjaer, 1996; Kang and Lindgren, 1998; Nikkanen and Ruotsalainen, 2000), in natural populations (Bila and Lindgren, 1998), and in plantation areas (Bila *et al.*, 1999). Research on cone and seed properties, phenology, and pollen production in seed orchards have also

been studied (Yazdani and Lindgren, 1991; Matziris, 1993, 1997, 1998; Fries, 1994; Yazdani et al., 1995; Keskin, 1999). Such fertility variation will cause rapid accumulation of group coancestry and potential inbreeding in the following generation (Bila, 2000; Kang, 2001). To monitor and determine the genetic diversity of seed crops, some genetic parameters such as genetic relatedness, inbreeding, and gene diversity should be calculated.

Effective population size and status number have been widely used to express accumulated inbreeding, random genetic drift and group coancestry (Crow and Kimura, 1970; Lindgren et al., 1996; Kjaer and Wellendorf, 1997). A key difference between effective population size and status number is how to describe the loss of gene diversity (inbreeding and group-coancestry). The former expresses it as dynamic over generations, while the latter does as static in each generation separately (Lindgren and Mullin, 1998). Seed orchards are generally used only for one generation. So, the status number is a good tool for monitoring gene diversity of orchard crops.

The purpose of this study is to evaluate fertility variation among orchard parents and its effect on gene diversity of seed (i.e., status number), and to provide genetic information to guide the "National Tree Breeding and Seed Production Programme for Turkey (Koski and Antola, 1993)". New seed orchards in Turkey will be established vegetatively with grafts after progeny tests according to the programme.

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Materials and Methods

Descriptions of seed orchards

The study was carried out in two clonal seed orchards of P. brutia and one clonal orchard of P. nigra and P. sylvestris, respectively. The seed orchards of P. brutia originated from plus trees, which were selected from seed stands at Bucak (orchard 1) and Serik (orchard 2), were established in 1985, at Antalya (latitude 37°20'N, longitude 30°35'E, altitude 310 m), southern part of Turkey. Grafts were planted at spacing 8 m x 8 m, and the seed orchards 1 and 2 are composing of 30 and 35 clones (1,620 and 1,080 ramets), respectively. The seed orchard of P. nigra originated from a plus tree stand at Tavsanli and that of P. sylvestris originated from a plus tree stand at Dirgine were established in 1986, at Adapazari (latitude 40°55' N, longitude 30°45' E, altitude 120 m), northern part of Turkey. Grafts were planted at spacing 8 m x 8 m in the P. nigra seed orchard consisted of 31 clones (1,340 ramets), and 7 m x 7 m in the P. sylvestris seed orchard consisted of 29 clones (1,265 ramets), respectively (Anonymous, 2001).

Data collection and parental-balance curve

The numbers of female and male strobili were counted from six ramets chosen randomly from each clone in each orchard. Strobili of the sampled grafts were counted individually over the whole crown. All clones in the studied seed orchard were surveyed at the beginning of April in 2001.

Parental-balance was assessed using a cumulative gamete contribution curve (GRIFFIN, 1982). The numbers of female and male strobili were ordered by clones from high to low strobilus production, and then the cumulative contribution percentages were plotted against the proportion of the clones.

Status number

We used the concept of status number $(N_s, {\rm LINDGREN}\ et\ al., 1996).$ If the clones are unrelated and non-inbred, the status numbers of female $(N_{s(f)})$ and male parents $(N_{s(m)})$ are calculated as

$$N_{s(f)} = \frac{1}{\sum_{i=1}^{N} p_{i(f)}^{2}}$$
 [1-a]

$$N_{s_{(m)}} = \frac{1}{\sum_{i=1}^{N} p_{i_{(m)}}^{2}}$$
 [1-b]

where $p_{i(f)}$ is the female fertility of the clone i, $p_{i(m)}$ is the male fertility of the clone i and N is the census number in the seed orchard. Here, fertility is estimated based on the strobilus assessment.

The central issue for describing fertility variation is the square sum of contribution as in formulae [1-a] and [1-b]. Kang and Lindgren (1998) defined and developed the sibling coefficient (Ψ) that expresses the fertility variation in a population using a coefficient of variation (CV) in strobilus production among clones, which is independent on the census number (N). Ψ is related to CV as $\Psi = N\Sigma p_i^2 = CV^2 + 1$. In principle, Ψ is regarded as a characteristic of the probability density function of fertility, while CV is a normalized observed variance that is dependent on the number of observation (Kang and Lindgren, 1998). When the total fertility (i.e., average of female and male fertility) is estimated, the correlation between female and male fertility is also important (Kang and El-Kassaby, 2002).

Status number on total fertility was calculated following $\ensuremath{\mathsf{KANG}}\xspace\,(2001)$ and as:

$$N_s = \frac{4N}{[\Psi_f + \Psi_m + 2 + 2r\sqrt{(\Psi_f - 1)(\Psi_m - 1)}]}$$
 [2]

where Ψ_f and Ψ_m are the fertility variation of female and male parents, which are equivalent to CV_f^2+1 and CV_m^2+1 , respectively (Kang, 2001). Here, CV_f and CV_m are the coefficients of variation in female and male fertility, and r is the correlation coefficient between female and male fertility.

Results and Discussion

Female and male strobilus production

The average, coefficient of variation (CV), range of female and male strobilus production and phenotypic correlation coefficients between female and male strobili are presented in *Table 1*.

Table 1. – Average, coefficient of variation (CV), range and correlation (r) in female and male strobilus production in the studied seed orchards in Turkey.

	Pinus brutia1		Pinus brutia 2		Pinus nigra		Pinus sylvestris	
	female	male	female	male	female	male	female	male
Average	89.0	289.8	93.6	317.0	78.3	187.4	201.4	531.3
CV	0.491	1.277	0.493	1.036	0.247	0.350	0,375	0.257
Range	0-368	0-2472	0-333	0-2720	14-210	0-600	0-910	136- 1768
r a)	0.278		0.701		0.387		0.774	

 a) phenotypic correlation coefficient between clonal female and male strobilus production

As shown in *Table 1*, there was a large difference in the production of female and male strobili among clones. The fertility variation (*CV*) in *Pinus brutia* was larger than *P. nigra* and *P. sylvestris*. Variation in male fertility was higher than in female fertility in *P. brutia* and *P. nigra* seed orchards. Also, the most abundant quartile of clones in *P. brutia* (orchards 1 and 2), *P. nigra* and *P. sylvestris* seed orchards produced 44.1%, 40.5%, 33.9% and 37.8% of total female, and 76.1%, 62.6%, 36.7% and 34.1% of total male strobilus production, respectively. Positive correlations between female and male fertility were found in all seed orchards (*Table 1*).

Parental-balance curves are shown in *Figure 1* by means of cumulative gamete contribution. Cumulative contribution of female and male parents showed that male fertility variation was very large in *P. brutia* seed orchards, compared to the other species. In the seed orchard of *P. britua* 1, six clones (20%) did not produce any male strobili, meaning that those clones do not contribute to gamete production as fathers. The curves in *P. nigra* and *P. sylvestris* seed orchards were less distorted than in *P. brutia*, implying that clones in *P. nigra* and *P. sylvestris* seed orchards contribute more equally to gamete production (Kang, 2000).

The *P. brutia* seed orchards were 16 years old at the time of the study, which is rather young in age to peak strobilus production. It is reported that fertility variation is higher when seed orchards are young (Kang and Lindgren, 1999; Bila, 2000). In the present study, data on strobilus production were collected from only one year. Kang (2001) reported that clonal fertility was fluctuated over many years in conifer seed orchards. Therefore, it is needed to collect more data on fertility variation to draw accurate conclusion. It should be emphasised in this study that there is the immediate need for fertility

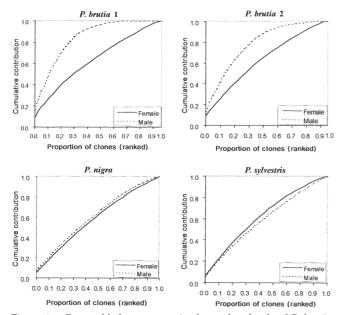


Figure 1. – Parental-balance curves in the seed orchards of *P. brutia*, *P. nigra* and *P. sylvestris* in Turkey. Proportion of clones was decreasingly ranked according to strobilus production.

information on young seed orchards to adjust orchard management options.

Flowering data were collected from six randomly selected ramets per clone. Many of the first-generation clonal seed orchards are established with intention to have nearly equal number of ramets for each clone. However, there could be ramet variation among clones because intentional (e.g., roguing) and unintentional (e.g., graft availability, graft-incompatibility) variations in ramet numbers among clones occur in the seed orchards (KANG et al., 2001). We did not consider pollen contamination in the present study. Pollen contamination is a main source of gene migration into seed orchard, which affects genetic quality of orchard crops (KANG, 2001). While ramet variation will decrease status number, gene migration from trees outside of the orchards will increase gene diversity (LIND-GREN and MULLIN, 1998).

 $Fertility\ variation,\ status\ number\ and\ relative\ status\ number$

Fertility variation, status number and relative status number for female and male strobili are presented in *Table 2*, and those factors for total fertility are shown in *Table 3*.

 $\begin{array}{l} \textit{Table 2.} - \textit{Fertility variation} \; (\Psi_f \& \, \Psi_m), \textit{status number} \; (N_{s(f)} \& \, N_{s(m)}) \; \textit{and} \\ \textit{relative status number} \; (N_{r(f)} \& \, N_{r(m)}) \; \textit{in gametic gene pool for female} \\ \textit{and male parents.} \end{array}$

	P. brutia 1 (30) a)		P. brutia 2 (35)		P. nigra (31)		P. sylvestris (28)	
	female	Male	female	male	female	Male	female	male
$\Psi_f \& \Psi_m^{\ b)}$	1.24	2.63	1.24	2.07	1.06	1.12	1.14	1.07
$N_{s(f)} & N_{s(m)}$	24.2	11.4	28.2	16.9	29.2	27.6	24.5	26.3
$N_{r(f)} & N_{r(m)}$	0.81	0.38	0.80	0.48	0.94	0.89	0.88	0.94

a) census number of clones (N) in parentheses

b) f and m represent female and male, respectively.

The status number and relative status number of female parents were higher than those of male parents in most seed orchards, except only in *P. sylvestris* (*Table 2*). The relative status numbers for male strobili of *Pinus brutia* (orchards 1 and 2) were very low, implying that some management options (*e.g.*, supplemental mass pollination) should be applied.

Fertility variation (Ψ) in P. brutia seed orchards was higher than that in P. nigra and P. sylvestris seed orchard (Table 3). Status number (N_s) for the total fertility was moderately high in all seed orchards (Table 3). But the relative status number (N_r) was low in P. brutia seed orchards, indicating that orchard may not be functioning efficiently. Relative gene diversity (GD) was maintained high (Table 3). Gene diversity of orchard crop can be estimated relative to the reference population where trees are unrelated and non-inbred (Kang and Lindgren, 1999). However, if the parents are related or inbred, GD in the studied seed orchard should be lower.

Table 3. – Fertility variation (Ψ) , status number (N_s) , relative status number (N_r) and relative gene diversity (GD) in total gametic gene pool in the studied seed orchards.

	P. brutia 1 (30) a)	P. brutia 2 (35)	P. nigra (31)	P. sylvestris (28)
Ψ	1.55	1.51	1.06	1.09
N_s	19.3	23.2	29.2	25.7
N_r	0.64	0.66	0.94	0.92
$GD^{b)}$	0.974	0.978	0.983	0.981

a) census number of clones (N) in parentheses

b) $GD = 1 - 1/(2*N_s)$

Based on genetic information from progeny tests, new seed orchards will be established in Turkey according the "Turkish Breeding Programme". Our results show that there is lack of pollen production in *Pinus brutia* seed orchard, which may cause to low seed production. Therefore, special attention should focus on the increase of pollen production in future orchards of this species. In the seed orchard of *P. sylvestris*, female fertility variation was higher than male fertility variation. So, equal seed harvest or mixing seed equally from clones may improve the female fertility variation and thus status number as suggested by KANG (2001).

Fertility variation may be controlled somewhat by ramet number when new seed orchards are established or existing orchards are rogued (Hodge and White, 1993; Kang et al., 2001). In advanced-generation seed orchards, the main criteria for the selection of orchard parents will be the genetic values of the candidates. However, fertility variation should also be considered to increase status effective number, especially in *P. brutia* seed orchards.

Turkish seed orchards have been established with about 30 clones originated from plus trees from a single seed stand (Anonymous, 2001). The genetic variation is therefore quite narrow, especially if orchard parents are related or somewhat inbred. The existing seed orchards do not have enough clones for genetic roguing based on progeny-test results in the future. Thus, the existing orchards should be managed by refreshment or infusion of new genetic materials. New orchards should also be established with more clones, which are genetically unrelated each other. Based on the Turkish breeding programme, the new seed orchards would be established in *P. brutia*, *P. nigra*, and *P. sylvestris*. In the establishment of new seed orchards or management of existing orchards, the effective number (e.g.,

status number) rather than census number should be addressed, considering relatedness and fertility variation, for the monitoring of gene diversity in orchard crops.

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References

Anonymous: 2000 yili calisma raporu, 2001 yili calisma programi (Working report of 2000 and working plan of 2001). The Research Directorate of Forest Tree Seeds and Tree Breeding, No. 3 (132/7), Ankara, Turkey (2001). — BILA, A. D.: Fertility variation and its effects on gene diversity in forest tree populations. Ph. D Thesis. Swedish University of Agricultural Science, Umeå, Sweden. Acta Universitatis Agriculturae Sueciae, Silvestria 166 (2000). — BILA, A. D. and LINDGREN, D.: Fertility variation in Milletia stuhlmanii, Brachystegia spiciliformis, Brachystegia bohemii and Leucaena leucocephala and its effects on relatedness in seeds. For. Genet. 5: 119-120 (1998). - BILA, A. D., LINDGREN, D. and MULLIN, T. J.: Fertility variation and its effect on diversity over generations in a Teak plantation (Tectona grandis L.f.). Silvae Genet. 48: 109-114 (1999). — CALISKAN, T.: Hizli gelisen turlerle ilgili rapor (Report of fast growing species in Turkey). Workshop-Hizli Gelisen Turlerle Yapilan Agaclandirma Calismalarinin Degerlendirilmesi ve Yapilacak Calismalar, 8-9 Aralik, Ankara. (1998). — CROW, J. F. and KIMURA, M.: An introduction to population genetics theory. Harper and Row Publishers, London, pp. 591 (1970). — FRIES, A.: Development of flowering and the effect of pruning in a clonal seed orchard of lodgepole pine. Can. J. For. Res. 24: 71-76 (1994). — GRIFFIN, A. R.: Clonal variation in radiata pine seed orchards. I. Some flowering, cone, and seed production traits. Aust. For. Res. 12: 295-302 (1982). - HODGE, G. R. and WHITE, T. L.: Advanced-generation wind-pollination seed orchard design. New Forests 7: 213-236 (1993). - KANG, K. S.: Clonal and annual variation of flower production and composition of gamete gene pool in a clonal seed orchard of Pinus densiflora. Can. J. For. Res. 30: 1275–1280 (2000). — KANG, K. S.: Genetic gain and gene diversity of seed orchard crops. Ph. D Thesis. Swedish University of Agricultural Science, Umeå, Sweden. Acta Universitatis Agriculturae Sueciae, Silvestria 187. pp. 75 (2001). — Kang,

K. S. and EL-KASSABY, Y. A.: Considerations of correlated fertility between genders on genetic diversity: Pinus densiflora seed orchard as a model. Theor Appl. Genet. 000-000 (in press) (2002). — KANG, K. S. and LINDGREN, D.: Fertility variation and its effect on the relatedness of seeds in Pinus densiflora, Pinus thunbergii and Pinus koraiensis clonal seed orchards. Silvae Genet. 47: 196-201 (1998). - KANG, K. S. and LINDGREN, D.: Fertility variation among clones of Korean pine (Pinus koraiensis S. et Z.) and its implications on seed orchard management. For. Genet. 6: 191-200 (1999). — Kang, K. S., Harju, A. M., Lindgren, D., Nikkanen, T., ALMQVIST, C. and SUH, G. U.: Variation in effective number of clones in seed orchards. New Forests 21: 17-33 (2001). - Keskin, S.: Clonal variation in flowering and cone characteristics in a Pinus brutia seed orchard. SAFRI Technical Bulletin No. 9, Antalya, pp. 96 (1999). -KJAER, E. D.: Estimation of effective population number in a Picea abies (KARST.) seed orchard based on flower assessment. Scand. J. For. Res. 11: 111-121 (1996). - KJAER, E. D. and WELLENDORF, H.: Variation in flowering and reproductive success in a Danish Picea abies (Karst) seed orchard. For. Genet. 4: 181-188 (1997). - Koski, V. and Antola, J.: National Tree Breeding and Seed Production Programme for Turkey 1994-2003. The Research Directorate of Forest Tree Seeds and Tree Breeding, Ankara, Turkey. (1993). - LINDGREN, D., GEA, L. and JEFFER-SON, P.: Loss of genetic diversity monitored by status number. Silvae Genet. 45: 52-59 (1996). — LINDGREN, D. and MULLIN, T. J.: Relatedness and status number in seed orchard crops. Can. J. For. Res. 28: 276-283 (1998). — MATZIRIS, D.: Variation in cone production in a clonal seed orchard of Black pine. Silvae Genet. 42: 136–141 (1993). — MATZIRIS, D.: Variation in growth, flowering and cone production in a clonal seed orchard of Aleppo pine grown in Greece. Silvae Genet. 46: 224–228 (1997). — MATZIRIS, D.: Genetic variation in cone and seed characteristics in a clonal seed orchard of Aleppo pine grown in Greece. Silvae Genet. 47: 37-41 (1998). — NIKKANEN, T. and RUOTSALAINEN, S.: Variation in flowering abundance and its impact on the genetic diversity of the seed crop in a Norway spruce seed orchard. Silva Fenn. 34: 205-222 (2000). — SAVOLINEN, O. and KÄRKKÄINEN, K.: Effect of forest management on gene pools. New Forests 6: 326-345 (1992). — YAZDANI, R. and LINDGREN, D.: Variation of pollen contamination in a Scots pine seed orchard. Silvae Genet. 40: 243-246 (1991). — YAZDANI, R., LINDGREN, D., YAZDANI, F. S., PASCUAL, L. and ERIKSSON, U.: Flowering, phenology, empty seeds and pollen contamination in a clonal seed orchard of Pinus sylvestris in northern Sweden. In: Population genetics and genetic conservation of forest trees. SPB Academic Publishing, Amsterdam, p. 309-319 (1995).

Inbreeding in Pinus Radiata

III. The Effect of Inbreeding on Age-age Correlation and Early Selection Efficiency

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Abstract

A breeding strategy involving inbreeding followed by cross-breeding of inbreds requires that the production of superior inbred lines must be possible, but crosses between lines should exhibit heterosis, inbreeding should not substantially delay reproduction, and early selection between lines to be effective. Age-age correlation and the effectiveness of early selection have been extensively reported for outcrossed populations of different species, but there are no reports for inbred populations. In this study, age-age correlations based on both family means and individual trees were investigated and compared in radiata pine populations with five different inbreeding levels $(F=0,\ 0.125,\ 0.25,\ 0.5$ and 0.75). Trends in additive genetic variance, environmental variance, heritability and age-age

additive genetic correlations were estimated from an outcrossed population (F=0).

For cross-sectional area at breast height, additive genetic variance increased from 3.7% at age 3 to 29.4% at age 5, remained at about 30% up to age 10, then declined to 15.6% at age 13.

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