

*altissima* SWINGLE. Abstracts VIIth International Congress on Plant Tissue Cell Culture, Amsterdam, June 24–29, Abstr. A3–161 (1990). — PIERIK, R. L. M.: In Vitro Culture of Higher Plants. Martinus Nijhoff, Dordrecht (1987). — QUOIRIN, M. and LEPOIVRE, Ph.: Improved medium for in vitro culture of *Prunus* sp. Acta Horticulturae 789, 437–442 (1977). — RAO, A. N., SIN, Y., KOTHAGODA, N. and HUTCHINSON, J. F.: Cotyledon tissue culture of some tropical fruits. pp. 124–137. In: Tissue Culture of Economically Important Plants. COSTED and ANBS National University. Ed. RAO, A. N., Singapore (1982). — SANKARA RAO, K.: Plantlets from somatic callus tissues of the East Indian Rosewood (*Dalbergia*

*latifolia* ROXB.). Plant Cell Reports 3, 199–201 (1986). — SANKARA RAO, K.: In vitro meristem cloning of *Eucalyptus tereticornis* Sm. Plant Cell Reports 7, 546–549 (1988). — SNEDECOR, G. W. and COCHRAN, W. G.: Statistical Methods Applied to Experiments in Agriculture and Biology. Allied Pacific Private Limited, Bombay (1956). — VARMA, R. V.: Seasonal incidence and possible control of important insect pests in plantations of *Allanthurus triphysa*. Kerala Forest Research Institute, Kerala (1986). — WELANDER, M.: In vitro rooting of the apple rootstock M26 in adult and juvenile growth phases and acclimatization of the plantlets. Physiologia Plantarum 58, 213–238 (1983).

## First Analysis on Allozyme Variation in Cedar Species (*Cedrus* sp.)<sup>1)2)</sup>

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### Summary

Isozyme variation among the four species of *Cedrus* genus was investigated in dormant vegetative buds of 25 trees of each species using starch gel electrophoresis. The most useful enzyme systems were LAP, MDH, 6PGD and PGI.

The results of this small study show that there is large heterozygosity in *Cedrus brevifolia*, *Cedrus libani* and *Cedrus atlantica*. In contrast, *Cedrus deodara* was fixed for the above studied enzymes. Moreover, it is shown that there are clear distinctions between *C. brevifolia*, *C. deodara* and the group of *C. atlantica-C. libani*.

**Key words:** *Cedrus atlantica*, *Cedrus brevifolia*, *Cedrus libani*, *Cedrus deodara*, isozyme variation, taxonomy.

### Introduction

The genus *Cedrus*, according to a number of authors, includes four coniferous evergreen tree species, with geographically separated distributions. *Cedrus brevifolia* HENRY in Cyprus, *Cedrus atlantica* MANETTI in Algeria and Morocco, *Cedrus libani* A. RICH in Lebanon, Syria and Turkey, *Cedrus deodara* LOUDON in Afganistan and India (M'HRIT, 1987; ARBEZ, 1987; DAVIS, 1965).

*Cedrus* has been successfully introduced in many countries outside of its natural distribution as ornamental and reforestation species. According to TOTH (1980) and M'HRIT (1987) it has, since the previous century, been introduced, to several European countries (France 1862, Italy 1866, Bulgaria 1890) and also into U. S. A. and Russia.

Due to the performance of its initial introductions, it soon became an important exotic species for Mediterranean and other countries with similar environmental conditions. In spite of the great interest in the genus, the limited information concerning the amount and pattern of its genetic variability, has been based mainly on provenance trials and studies of anatomical and morphological

traits (FAO, 1989). On the other hand, in the best of our knowledge, there has been no publication, on enzyme system studies in the genus cedar.

Our objective was to determine the level of genetic variability among and within species, as well as the usefulness of isozymes for taxonomic classification of the genus cedar.

### Material and Methods

#### Sample collection

Buds were collected from 25 randomly selected mature trees of each species and were stored in  $-20^{\circ}\text{C}$ . The material for *C. brevifolia* came from a natural stand of cedar in Cyprus (Paphos forest), whereas that for the other three species came from a species and provenance plantation established in 1968, in the arboretum of Loutra Thermis — Thessaloniki.

Seeds were also used in order to carry out a segregation analysis for those trees of *Cedrus brevifolia*, which proved to be heterozygous for at least one locus from the bud-isozyme analysis.

#### Electrophoresis procedure

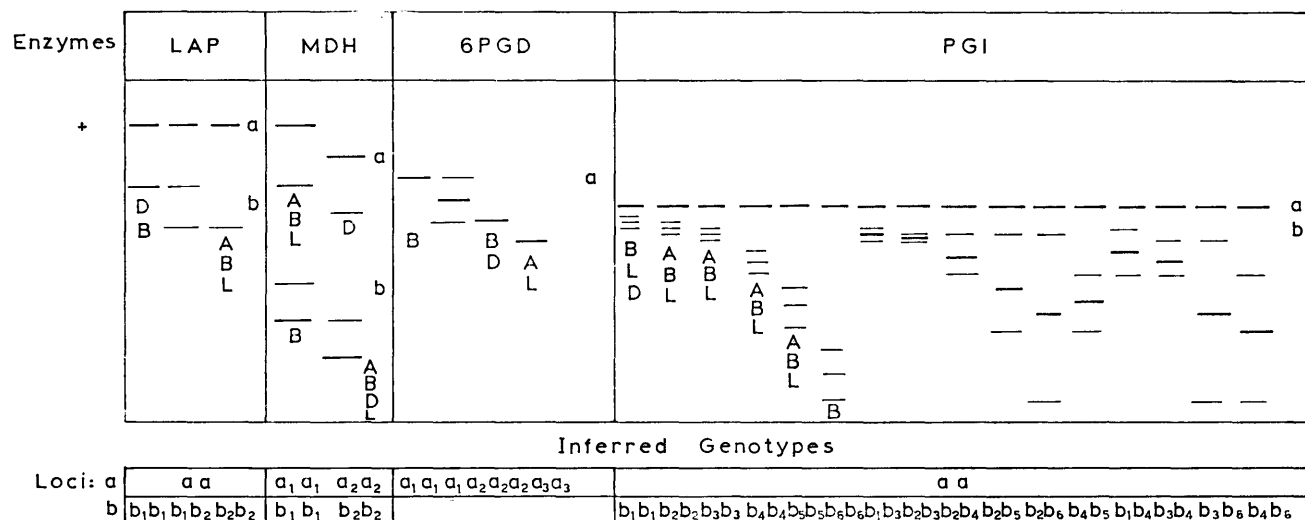
Four buds from each tree were homogenized with 0.3 ml vegetative extraction buffer pH 7 (CHELIAK and PITEL, 1984).

Megagametophytes dissected from germinated seeds (inoculated to germinate with cold stratification) with a radicle of 3 mm to 5 mm long, were homogenized separately in 0.4 ml seed extraction buffer, pH 7.6. The extraction buffer consisted of 0.1 M Tris, 3% (W/V) PVP-40, and 5 drops of b-mercaptoethanol. pH was adjusted to 7.6.

The homogenates were analyzed for: acid phosphatase (ACP E.C. 3.1.3.2.), aspartate aminotransferase (AAT or GOT E.C. 2.6.1.1.), diaphorase (DIA E.C. 1.6.4.3.), leucine aminopeptidase (LAP E.C. 3.4.11.1.), malate dehydrogenase (MDH E.C. 1.1.1.37.), peroxidase (PER E.C. 1.11.1.7.), 6-phosphogluconate dehydrogenase (6PGD E.C. 1.1.1.44.) and phosphoglucose isomerase (PGI E.C. 5.3.1.9.), in horizontal electrophoresis system with 11.5% (W/V) starch. Gels were prepared from a mixture (9:3 W/V) of MERK and CONNAUGHT starch.

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A: *Cedrus atlantica*  
 B: *Cedrus brevifolia*  
 D: *Cedrus deodara*  
 L: *Cedrus libani*

Figure 1. — Banding patterns observed in diploid tissues (buds) for four enzymes in *Cedrus* sp..

Composition of gels, electrode buffers and staining recipes were similar to those of CHELIAK and PITEL (1984), except for the staining recipe of MDH, which was similar to that of CONCLE et al. (1982).

The alleles in each locus (allozymes) and the loci (isozymes) were numbered in decreasing order of anodal mobility.

Estimation of species heterozygosity (a measure of the genetic variation within the species) and the "genetic distance" (a measure of the differentiation between species) was based on the observed gene frequencies and was estimated according to NEI (1977, 1978). Cluster analysis, using the unweighted pair group method, was performed on the matrix of NEI's genetic distance.

### Results

Six enzyme systems out of eight i. e. LAP, MDH, 6PGD, PGI, GOT, DIA, representing a number of 12 loci, were resolved with sufficient consistency and clarity. The isozyme banding patterns of LAP, MDH, 6PGD and PGI enzyme systems and the allele frequencies at each locus are shown in figure 1 and table 1 respectively. GOT and DIA enzyme systems appeared to be monomorphic in all species.

#### Leucine aminopeptidase (LAP)

Two zones of activity were observed for LAP which appeared to represent two distinct loci. The most anodal

zone (LAPa) appeared to be monomorphic in all species. The cathodal zone (LAPb) produced two variants (allele b<sub>1</sub>, b<sub>2</sub>). In *C. brevifolia*, both b<sub>1</sub> and b<sub>2</sub> were found, while in *C. deodara* only b<sub>1</sub>, and in *C. libani* and *C. atlantica* only b<sub>2</sub>, were detected.

#### Malate dehydrogenase (MDH)

Two zones of activity were also observed for MDH. In both zones (locus) double-banded allozymes were present.

In the first locus (MDHa) 2 variants (a<sub>1</sub>, a<sub>2</sub>) were scored. Allele a<sub>2</sub> was species-specific to *Cedrus deodara* while a<sub>1</sub> was found in homozygous state in the other species.

In the cathodal locus (MDHb) again two alleles were detected. Allele b<sub>2</sub> was found in all species, but b<sub>1</sub> was found only in *C. brevifolia*.

#### 6-Phosphogluconate dehydrogenase (6PGD)

6PGD behaved as a dimeric enzyme with a single locus and three variants (alleles a<sub>1</sub>, a<sub>2</sub>, a<sub>3</sub>). Allele a<sub>3</sub> was found only in *Cedrus atlantica* and *Cedrus libani* at frequencies of 100%.

#### Phosphoglucose isomerase (PGI)

Two zones of activity were observed for PGI. The most anodal zone (loci: PGIa) was not resolved satisfactorily, but seemed to be monomorphic. The second one PGIb, appeared as a triple band isozyme with six variants (alleles b<sub>1</sub>, b<sub>2</sub>, b<sub>3</sub>, b<sub>4</sub>, b<sub>5</sub> and b<sub>6</sub>). In the heterozygous phenotypes hybrid bands appeared, implying a possible dimeric

Table 1. — Allele frequencies of 5 isozyme loci in diploid tissue of *Cedrus* sp..

Enzymes and Alleles	LAP <sub>b</sub>		MDH <sub>a</sub>		MDH <sub>b</sub>		6PGD			PGI <sub>b</sub>					
	b <sub>1</sub>	b <sub>2</sub>	a <sub>1</sub>	a <sub>2</sub>	b <sub>1</sub>	b <sub>2</sub>	a <sub>1</sub>	a <sub>2</sub>	a <sub>3</sub>	b <sub>1</sub>	b <sub>2</sub>	b <sub>3</sub>	b <sub>4</sub>	b <sub>5</sub>	b <sub>6</sub>
<i>Cedrus atlantica</i>	-	1.00	1.00	-	-	1.00	-	-	1.00	-	0.20	0.16	0.38	0.26	-
<i>Cedrus brevifolia</i>	0.82	0.18	1.00	-	0.04	0.96	0.58	0.42	-	0.10	0.20	0.20	0.24	0.10	0.16
<i>Cedrus deodara</i>	1.00	-	-	1.00	-	1.00	-	1.00	-	1.00	-	-	-	-	-
<i>Cedrus libani</i>	-	1.00	1.00	-	-	1.00	-	-	1.00	0.10	0.46	0.20	0.20	0.04	-

Table 2. — Genetic variability of five loci in *Cedrus* diploid tissue.

Species	Number of trees	Number of alleles	Observed Heterozygosity ( $H_o$ )	Expected Heterozygosity ( $H_e$ )	Chi-square test ( $X^2$ )	Degrees of Freedom
<i>Cedrus atlantica</i>	25	8	0.1280 ± 0.128	0.1474 ± 0.147	0.0026	4
<i>Cedrus brevifolia</i>	25	13	0.2400 ± 0.115	0.3420 ± 0.151	0.0304	4
<i>Cedrus deodara</i>	25	5	0.0000 ± 0.000	0.0000 ± 0.000	0.0000	4
<i>Cedrus libani</i>	25	9	0.1440 ± 0.144	0.1422 ± 0.142	0.0000	4

Table 3. — Unbiased genetic distances between *Cedrus* sp..

	<i>C. atlantica</i>	<i>C. libani</i>	<i>C. deodara</i>
<i>C. brevifolia</i>	0.4749	0.4759	0.5672
<i>C. atlantica</i>		0.01554	1.5297
<i>C. libani</i>			1.4374

structure of the enzyme. Alleles  $b_1$ ,  $b_2$ ,  $b_3$ ,  $b_4$  and  $b_5$  were found in *C. libani* whereas  $b_2$ ,  $b_3$ ,  $b_4$  and  $b_5$  in *C. atlantica*. *C. deodara* had only one allele  $b_1$ , while *C. brevifolia* had all of them.  $b_6$  was species-specific to *C. brevifolia* with a frequency of 16%. The same isozyme banding patterns were found in both haploid and diploid tissues (megagametophytes) of *Cedrus brevifolia*, which were analysed to test Mendelian ratios. The Chi-square test ( $X^2 = 0.32 < X^2_{0.05,49}$ ) showed that the segregation fitted the expected Mendelian ratio of 1:1.

In the case of MDH in haploid tissue, an additional locus with very low mobility also appeared, while the active loci were those found in the diploid tissue. It seems that in the genus *Cedrus* only three loci of MDH are present, corresponding to MDH<sub>a</sub>, MDH<sub>b</sub> and MDH<sub>d</sub> (EL-KASSABY, 1981), of which two appeared in the diploid tissue and three in the haploid. The lack of MDH<sub>c</sub> locus and the heterodimeric band, which is the product of MDH<sub>b</sub> and MDH<sub>c</sub>, are lacking probably in *Cedrus*, as it was also detected in other conifers, e. g. *A. grandis*, *A. concolor* (EL-KASSABY, 1981).

Genetic variation for each species of *Cedrus* was expressed through the expected heterozygosities, which were 0.3420 for *C. brevifolia*, 0.1474 for *C. atlantica*, 0.1422 for *C. libani* and 0 for *C. deodara* (Table 2). The Chi-square test showed that there were no significant differences between observed and expected heterozygosities.

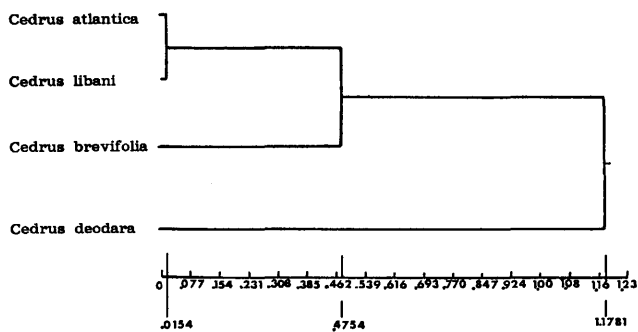


Figure 2. — Dendrogram of 4 *Cedrus* species, based on 5 polymorphic loci, constructed from the genetic distances estimated according to Nei's (1978) unbiased genetic distance.

The cluster analysis of unbiased genetic distances (Table 3, Figure 2) formed two distinct clusters: (a) *C. atlantica* and *C. libani*, with a very small genetic distance (0.0154), (b) *C. deodara*, with the largest genetic distance (0.1781). *C. brevifolia* occupied the place between the above 2 clusters but closer to the first one.

### Discussion

Isozyme analysis of *Cedrus* diploid tissue has provided new information about the geographic distribution and the relative amounts of genetic variation in the genus. As was mentioned in the introduction, there has been no report based on isozyme data concerning the genus *Cedrus* yet, whereas the existing information is mainly based on morphological and anatomical traits.

The data of isozyme analysis raises doubts about the separation of *C. atlantica* and *C. libani* into 2 distinct species, since no distinguishing gene marker was detected to justify their taxonomic status. On the contrary, they have allele  $a_3$  as a common gene marker in 6PGD, which distinguishes them from the other two species (*C. brevifolia* and *C. deodara*). The absence of allele  $b_1$  in PGIB in *C. atlantica* may be due to sampling error or to a possible over-estimate of the  $b_2$  allele, which was very close to  $b_1$  (Fig. 1). In contrast, for MDH<sub>a</sub>, allele  $a_2$  is a gene marker for *C. deodara* since it was not detected in the other species. *Cedrus brevifolia* has the  $b_6$  allele in PGIB as a gene marker, which was not detected in the other species.

Concerning the dimeric structure of PGI and 6PGD enzyme systems the findings of the present work are in agreement with those of NEALE and ADAMS (1981) for *Abies balsamea* and ADAMS and JOLY (1980) for Loblolly Pine, MITTON et al. (1979) for Ponderosa Pine, GURIES and LEDIG (1978) for *Pinus rigida*, SCALTSOYIANNES et al. (1990) for the Mediterranean species and provenances of *Abies*, NEALE et al. (1984), ADAMS et al. (1990) and EL-KASSABY et al. (1982) for Douglas fir. The multibanded pattern of PGIB allozymes was also noticed by ADAMS et al. (1990) in *Pseudotsuga menziesii* var. *menziesii* and *Scaltskyiannes* et al. (unpublished) in Moroccan firs (*A. pinsapo* var. *marocana* and var. *tazaotana*).

The monomeric subunit structure of LAP enzyme system is in agreement with the findings for other conifers (CONKLE, 1971; ADAMS and JOLY, 1980; MILLAR, 1985).

The dendrogram (Fig. 2), constructed from the genetic distances estimated (Table 3), implies that *C. atlantica* is similar to *C. libani*. In contrast, there is a clear distinction between those two and the group *C. brevifolia* and *C. deodara*. The low differentiation between *C. libani* and *C. atlantica* was expected, since the break of physical connection between the 2 so-called species, due to the

desertification of North Africa is relatively recent, so there was not enough time for species differentiation. In the Med-Checklist (GREUTER et al., 1984), *Cedrus atlantica* is listed as *Cedrus libani* subsp. *atlantica*, implying an uncertainty as to the existence of 2 separate species. The slight differentiation between those two species, could also be attributed to the origin of the material used in this study, which was collected from an arboretum where the trees may have been derived from only a few parental trees.

The heterozygosity (Table 2) shows that *C. brevifolia* has high variation (0.3420) while *C. deodara* has no variation at all. *C. libani* and *C. atlantica* are intermediate.

Generally, conifers exhibit high levels of heterozygosity. Up to now a notable exception to this rule was *Pinus resinosa* (FOWLER and MORRIS, 1977), probably as a result of a severe population restriction during the Pleistocene period. The fixation (zero heterozygosity) of *C. deodara* was unexpected, since the species occupies large areas, of about 500.000 ha (M'HIRIT, 1987) in Afganistan and the Himalayas. A possible explanation of this observation is the nature of the material used in the isozyme analysis. As noted above, samples of *C. deodara* were collected from an arboretum established with seeds of unknown origin. The lack of variation can be attributed to fixation of the donor source or the possible limited variation of the "European population".

Comparing the heterozygosity of the *Cedrus* species, excluding *C. deodara*, with that of other coniferous species, it appears that the heterozygosity of *C. brevifolia* is similar to that of *Abies* sp. (SCALTSOYIANNES, PANETSOS and ZARAGOTAS, 1990; SCALTSOYIANNES and PANETSOS, unpublished) while *Cedrus libani* and *Cedrus atlantica* have less heterozygosity.

This interpretation of enzyme variation of *Cedrus* sp. has important implications for future selection and improvement of the species. For *Cedrus deodara* more research is needed to solve the problem of its fixation. Research material should be derived from stands of its natural distribution. Further research on *C. atlantica* and *C. libani*, based on material from natural stands, will elucidate their taxonomic status.

Isozyme studies based on haploid tissue (endosperm) of *C. brevifolia* (unpublished) revealed the same pattern as the one presented in this study, coming from diploid tissue.

## References

- ADAMS, W. T. and JOLY, R. J.: Genetics of allozyme variants in loblolly pine. *J. Hered.* 71: 33-40 (1980). — ADAMS, W. T., NEALE, D. B., DOERKSEN, A. H. and SMITH, D. B.: Inheritance and Linkage of Isozyme Variants from seed and vegetative bud tissues in coastal Douglas-fir (*Pseudotsuga menziesii* var. *menziesii* (MIRB.) FRANCO). *Silvae Genetica* 39 (3-4): 153-167 (1990). — ARBEZ, M.: Les ressources genetiques forestieres en France. Les coniferes 1. INRA, Paris (1987). — BERGMANN, F.: Adaptive acid phosphatase polymorphism in conifer seeds. *Silvae Genetica* 24 (5-6): 175-177 (1975). — CHELIAK, W. M. and PITEL, J. A.: Techniques for starch gel electrophoresis of enzymes from forest tree species. Inf. Rep. PI-X-2, Petawawa Nat. For. Inst., Canadian For. Serv., Agric. Canada (1984). — CONKLE, M. T.: Inheritance of alcohol dehydrogenase and leucine aminopeptidase isozymes in knobcone pine. *For. Sci.* 17: 190-194 (1971). — CONKLE, M. T.: Isozyme variation and linkage in six conifer species. *Proc. Symp. on Isozymes of N. Am. Forest Trees and Forest Insects*, Berkeley, California (1979). — CONKLE, W. M., HODGSKISS, P. D., NUNNALLY, L. B. and HUNTER, S. C.: Starch gel electrophoresis of conifer seeds: a laboratory manual. USDA For. Serv. Gen. Tech. Rep. PSW-64, Pac. Southwest For. Range Exp. Stn. Berkeley, CA (1982). — DAVIS, P. H.: Flora of Turkey and East Aegean Islands. Vol. I. 71-72: Univ. Press, Edinburgh (1965). — EL-KASSABY, Y. A.: Genetic interpretation of malate dehydrogenase isozymes in some conifer species. *The Journal of Heredity* 72, 451-452 (1981). — EL-KASSABY, Y. A., YEH, F. C. and SZIKLAI, O.: Inheritance of allozyme variants in coastal Douglas fir (*Pseudotsuga menziesii* var. *menziesii*). *Can. J. Genet. Cytol.* 24: 325-335 (1982). — FAO: Silviculture of Species: *Cedrus* spp. "Silva Mediterranea". FO: SCM/89/6 (1989). — FOWLER, D. P. and MORRIS, R. W.: Genetic diversity in red pine: evidence for low genetic heterozygosity. *Can. J. Forest Res.* 7: 343-347 (1977). — GREUTER, W., BURDET, H. M. and LONG, G.: Med-Checklist. 1. Pteridophyta (ed. 2) Gymnospermae Dicotyledonas (Acanthaceae — Gneoraceae). Edition des Conservatoire et jardin botaniques de la ville de Genève (1984). — GURIES, R. P. and LEDIG, F. T.: Inheritance of some polymorphic isoenzymes in pitch pine (*Pinus rigida* MILL.). *Heredity* 20: 27-32 (1978). — M'HIRIT, O.: Etat actuel des connaissances sur le Cedre. Element. pour un programme de recherche. Rapport Comité C. F. A./C.E.F./C.F.P.O. "Silva Mediterranea" (1987). — MILLAR, C. I.: Inheritance of allozyme variants in Bishop pine (*Pinus muricata* D. DON.). *Biochem. Genet.* 23: 933-946 (1985). — MITTON, J. B., LINHART, Y. B., STURGEON, K. B. and HAMRICK, J. L.: Allozyme polymorphisms detected in mature needle tissue of ponderosa pine. *J. Hered.* 70: 86-89 (1979). — NEALE, D. B. and ADAMS, W. T.: Inheritance of isozyme variants in seed tissues of balsam fir (*Abies balsamea*). *Can. J. Bot.* 59: 1285-1291 (1981). — NEALE, D. B., WEBER, J. C. and ADAMS, W. T.: Inheritance of needle tissue isozymes in Douglas-fir. *Can. J. Genet. Cytol.* 26: 459-468 (1984). — NEI, M.: F-Statistics and analysis of gene diversity in subdivided populations. *Ann. Hum. Genet.* 41: 225-233 (1977). — NEI, M.: Estimation of average heterozygosity and genetic distance from a small number of individuals. *Genetics* 89: 583-590 (1978). — SCALTSOYIANNES, A., PANETSOS, K. P. and ZARAGOTAS, D.: Genetic variation of Greek-fir as determined by isozyme analysis and its relation to other Mediterranean Firs. *Publ. EEC Cat. No CD-NA-13491-2A-C*: 99-117 (1991). — SCALTSOYIANNES, A. and PANETSOS, K. P.: Isozyme variation of Mediterranean Firs. (unpublished). — TOTI, J.: Le Cedre dans les pays du pourtour Mediterranee et dans deux autres pays et son importance forestiere. *Forests Mediterraneees* II, (1), 23-30 (1980).

## Inheritance and Linkage of Some Allozymes in *Taxus baccata* L.

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### Summary

Eleven enzyme systems coding for 22 loci were assayed in *Taxus baccata*. Mendelian inheritance was confirmed

for allozymes at 11 loci by testing the fit of band-pattern segregation in macrogametophytes from heterozygous trees to the expected 1:1 ratio.