

5. Conclusions

Somewhat changed silvicultural methods may be needed when using grafted clones in forestry. Choice of proper spacing is then important and research on the effect of competition and on the method at plant propagation is needed. On the whole the variation between plants within the seedling material was larger than between grafts within the clones except for straightness (*Figure 3 a—g*). The reason could be that genetic differences within the seedling material caused more variation than the genetic differences in the root part and the physiological differences resulting from the grafting procedure.

It seems to be possible to decrease variation, especially for volume production, by conducting trials with grafted clones, but then competition must be low. Clones could thus be a great help in breeding work. A greater number of clones must be tested to give information of interaction between genotype and spacing.

6. Acknowledgements

I am grateful to VERNER ÅKEBRAND, who took lead of the measurements with me as an assistant.

My thanks are also due to my supervisor DAG LINDGREN and to OLA LINDGREN, Department of Biometry and Forest Management, Umeå, for help during the statistical treatment of the material, and to all the other personnel at the Department of Forest Genetics and Plant Physiology, Umeå, who have helped me in different ways during my work on this paper.

7. Literature Cited

ANDERSSON, E. and HATTEMER, H. H.: Variation among Clones and Ortet-Ramet Relationship in Grafted Scots Pine (*Pinus sylvestris* L.). *Studia Forestalia Suecica* 148: 1—32 (1978). — ANDERSSON, S.-O.: Rönjningsförbandets betydelse för framtida gagnvirkesproduktion och kvalitet. Några försöksresultat och synpunkter. Föredrag hösten 1972. Royal College of Forestry, Stockholm: 1—40, (1973). —

BLOMQVIST, S.: Samband plusträd — kloner — avkomor av tall. Föreningen skogsträdsförädling. Institutet för skogsförbättring, 1975, årsbok: 171—194. Föreningen skogsträdsförädling. Institutet för skogsförbättring, 1st ed. 194 pp. 1976, Uppsala, Sweden (1976). — ELFVING, B.: Volume and structure in unthinned stands of Scots pine. Department of Forest Yield Research. Research Notes No. 35: 1—128, Royal College of Forestry, 1st ed, 128 pp. 1975, Stockholm (1975). — FRIES, A.: Genotyp-miljösamspel i ett tallympförsök. Arbetsrapport nr 3, Institutionen för skoglig genetik och växtfysiologi, Skogshögskolan, Sveriges Lantbruksuniversitet, Umeå, Sweden: 1—59 (1982). — HATTEMER, H. H., ANDERSSON, E. and TAMM, C.-O.: Effects of Spacing and Fertilization on Four Grafted Clones of Scots Pine. *Studia Forestalia Suecica* 141: 1—32 (1977). — HELLSTRÖM, C.: Reagerar talkloner olika på förband och gödsling? — Examensarbete i skogsgenetik. Arbetsrapport nr 1, Institutionen för skoglig genetik och växtfysiologi, Skogshögskolan, Sveriges Lantbruksuniversitet, Umeå: 1—63 (1982). — NETER, J. and WASSERMAN, W.: Applied Linear Statistical Models. Regression Analysis of Variance, and Experimental Designs. Irwin-Dorsey Limited, Georgetown, Ontario: 468—470, 1st ed. 863 pp. Georgetown, Ontario (1974). — NÄSLUND, M.: Functions and tables for computing the cubic volume of standing trees. Pine, spruce and birch in southern Sweden, and in the whole of Sweden (1947). — Reports of the Forest Research Institute of Sweden. Vol. 36: 1—81. Stockholm (1948). — PERSSON, A.: The influence of spacing on the quality of sawn timber from Scots pine. Department of Forest Yield Research. Research Notes No. 42: 1—128, Royal College of Forestry, 1st ed, 128 pp. Stockholm (1976). — PERSSON, A.: Quality development in young spacing trials with Scots pine. Department of Forest Yield Research. Research Notes No. 45: 1—152, Royal College of Forestry, 1st ed, 152 pp. Stockholm (1977). — ROULUND, H.: Growth and quality characters, their variation and correlation in a combined clone and progeny experiment in Norway spruce (*Picea abies* L. KARST). *Forest Tree Improvement* 14: 1—48, (1980). — SAKAI, K.-I. and MUKAIDE, H.: Estimation of Genetic, Environmental and Competitive Variances in Standing Forests. *Silvae Genetica* 16: 149—152 (1967). — VON EULER, F.: Kvalitet i ett klonförsök med tall. Examensarbete i skogsgenetik. Intern rapport nr 46, Institutionen för skoglig genetik och växtfysiologi, Skogshögskolan, Sveriges Lantbruksuniversitet Umeå: 1—34 (1982). — WELLENDOFF, H.: Resemblance in height growth between original trees and clones of Scots pine (*Pinus sylvestris* L.) *Forest Tree Improvement* 1: 25—45. Akademisk Forlag, Kobenhavn (1970).

The somatic chromosomes of *Cryptomeria japonica* with special reference to the marker chromosomes¹⁾

By S. E. SCHLARBAUM, T. TSUCHIYA and L. C. JOHNSON²⁾

(Received 2nd September 1983)

Summary

The diploid chromosome number of *Cryptomeria japonica* was found to be $2n = 2x = 22$, which concurs with previous reports. The majority of chromosomes were found to have median kinetochores. One submetacentric chromosome pair has an unusually long primary constriction which consists of chromomeres connected by the kinetonema. The possibility that this unusual region is of a com-

pound nature, containing the nucleolar organizer and microtubule attachment sites, is discussed. Observations of the present study were compared with results of previous studies in order to provide a better understanding of the chromosome morphology in *Cryptomeria*.

Key words: *Cryptomeria*, *Taxodiaceae*, marker chromosomes, kinetochore, kinetonema.

Zusammenfassung

Die diploide Chromosomenzahl von *Cryptomeria japonica* hat sich als $2n = 2x = 22$ herausgestellt, was mit früheren Berichten übereinstimmt. Die Mehrzahl der Chromosomen zeigten mittelgroße Kinetochoren. Ein submetazentrisches Chromosomenpaar hat eine ungewöhnlich lange primäre Konstriktion, die aus Chromomeren besteht, welche durch Kinetonemen verbunden sind. Die Möglichkeit, daß diese ungewöhnliche Region zusammengesetzter Natur ist und einen nukleolaren Organisator und mikrotubulare Anknüpfungsstellen enthält, wird diskutiert. Beobachtungen der gegenwärtigen Untersuchung wurden mit den Ergeb-

¹⁾ Contributed from Department of Agronomy and published with the approval of the Director of the Colorado State University Experiment Station as Scientific Series Paper No. 2809.

²⁾ Former Graduate Research Assistant, and Professor, Department of Agronomy, Colorado State University, Ft. Collins, Colorado 80523, USA; and Northeastern Area Geneticist, State and Private Forestry, Forest Service — USDA, 1992 Folwell Ave., St. Paul, Minnesota 55108, USA. Dr. SCHLARBAUM is presently Assistant Professor, Dept. of Forestry, Wildlife and Fisheries, University of Tennessee, Knoxville, T N 3790.

Correspondence address: Dr. T. TSUCHIYA, Department of Agronomy, Colorado State University, Ft. Collins, CO 80523, USA.

nissen früherer Untersuchungen verglichen, um ein besseres Verständnis der Chromosomen-Morphologie bei *Cryptomeria* zu erlangen.

Introduction

Cryptomeria japonica D. DON, Japanese Sugi, is a monotypic genus in the redwood family, *Taxodiaceae*. The conifer is widely planted throughout the world but thought to be endemic only to Japan since the Pliocene era (FLORIN, 1963). Morphologically, *Cryptomeria* is distinct within *Taxodiaceae*. The genus is invariably classified in a monotypic tribe (HIDA, 1962; ECKENWALDER, 1976) or even separated from *Taxodiaceae* into a monotypic family (HAYATA, 1932).

The chromosome number of *Cryptomeria*, $2n = 2x = 22$, was initially reported by SAX and SAX (1933) and MATSUMOTO (1933) in independent studies. Chromosome counts in later studies have confirmed the initial observations (ZINNAI, 1947; KANAZAWA, 1949; CHIBA, 1950; ZINNAI and CHIBA, 1952; SAITO and HASHIZUME, 1958; SHIBATA *et al.*, 1959; MATSUDA and MIYAJIMA, 1976; SCHLARBAUM and TSUCHIYA, 1981). Chromosome morphology was studied in detail by SAX and SAX (1933), MATSUMOTO (1933), STIFF (1952), MEHRA and KHOSHOO (1956), KUROKI (1969), TODA (1977, 1979 a, b, 1980, 1981 a, b), MEHRA and ANAND (1979) and SOMEGO and KIKUTI (1980). However, descriptions of the *Cryptomeria* karyotype are conflicting among these previous studies. The present authors, therefore, undertook a study of the somatic chromosomes of *Cryptomeria* to make accurate observations on the chromosome complement and discuss these observations in relation to previous studies. Additionally, the study was undertaken to investigate for the presence of marker chromosomes with unusual structures which have been previously reported to occur in taxodiaceous tree species (SCHLARBAUM and TSUCHIYA, 1975, 1981, 1984; SCHLARBAUM, 1980; SCHLARBAUM *et al.*, 1983).

Materials and Methods

A single clone was selected for study from three *Cryptomeria* populations, respectively, located in Kyushu, Aomori Prefecture and Kumamoto Prefecture. Somatic cells in the root tip meristematic tissue were isolated and analyzed.

The root tips were immersed in 8-hydroxyquinoline (0.002 M) for 24–36 hours at 4° C and fixed in a 3:1 mix-

ture of ethyl alcohol and glacial acetic acid. After several days in the fixative, the root tips were hydrolyzed in 1 N HCl at 60° C for 10–15 minutes and stained with Feulgen or acetocarmine (SAYLOR, 1961; Schlarbaum and Tsuchiya, 1976).

The squash technique was used for slide preparation. Slides were made permanent by applying several drops of a 10:1 mixture of 45 percent acetic acid and glycerol to the edge of the cover slip (TSUCHIYA, 1971).

Results

The diploid chromosome number of *Cryptomeria* was observed as $2n = 22$ (Fig. 1) as previously reported by SAX and SAX (1933), MATSUMOTO (1933), and in other studies. Numerous cells (ca. 20–40 from each clone source) were present in which the diploid number was confirmed and the morphological features of the chromosomes were easily observed. There were no notable differences in chromosome morphology among the clones from the three populations.

With few exceptions, all of the chromosomes were metacentric (Fig. 1). However, one chromosome pair was submetacentric and exhibited a long kinetochore region. As clearly shown at meta-anaphase (Fig. 2 a) and metaphase (Fig. 2 b), this region was not an artifact nor an aberration but a consistent feature of the chromosome complement. The length of this kinetochore region varied in different cells at the same stage of mitosis and some variation was observed between homologous chromosomes in the same cell which may be partly ascribed to artifacts (Fig. 2 b). The region apparently consists of chromomeres connected by a darkly stained chromonema or kintonema (MATSUURA, 1941). In some slide preparations, the kinetochore region showed a spiral structure (Fig. 2 b - third chromosome from left). Occasionally, two large chromomeres were observed near the middle of the region as shown by several metaphase chromosomes in Figure 2 b. Conventional satellited chromosomes were not observed in the chromosome complement of *Cryptomeria*.

The chromosome complement of *Cryptomeria*, with the exception of the marker chromosome pair, was generally symmetrical in kinetochore position and relative size.

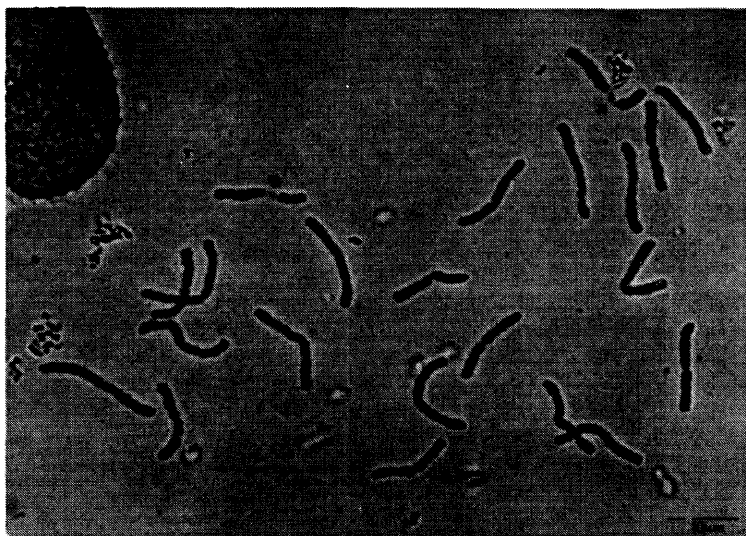


Figure 1. — Somatic metaphase cell of *Cryptomeria japonica* ($2n = 22$). Arrows indicate marker chromosomes with long kinetochore regions.

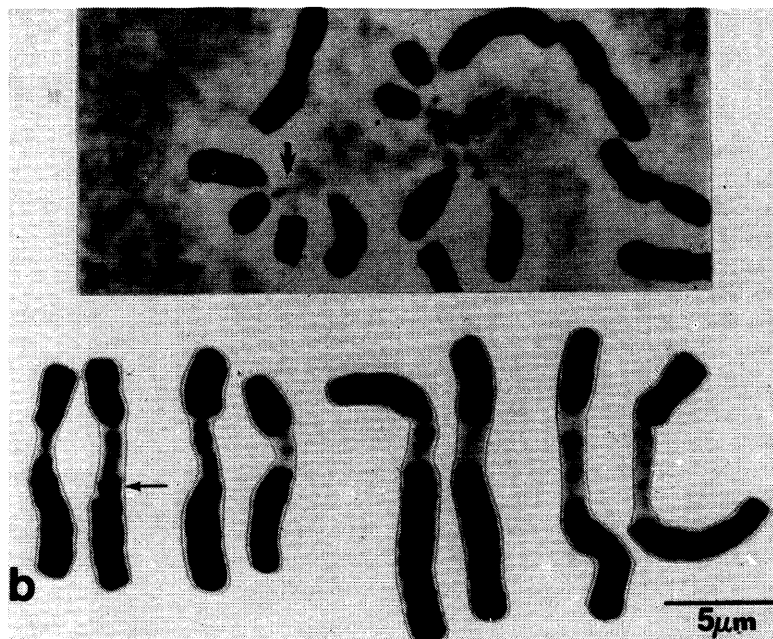


Figure 2. — a) Marker chromosome pair of *Cryptomeria japonica* at meta-anaphase. Arrows indicate long kinetochore regions. b) Isolated marker chromosome pairs at metaphase from four cells of *Cryptomeria japonica*. The "Köpfchen" is indicated by an arrow in the right chromosome of the first pair. The spiralization of the kintonema is evident in the left chromosome of the second pair.

Discussion

The symmetrical chromosome complement of *Cryptomeria* described in the present study concurs with previous investigations reported by KUROKI (1969), MEHRA and KHOSHOO (1956), TODA (1977, 1979 a, b, 1980, 1981 a, b), MEHRA and ANAND (1979) and SOMEGO and KIKUTI (1980). The studies of SAX and SAX (1933) and STIFF (1952) indicate conflicting viewpoints, reporting the presence of many nonmetacentric chromosomes in the complement. However, neither SAX and SAX (1933) nor STIFF (1952) presented a photograph of the chromosomes nor recognized the presence of chromosomes with unusual features. These two studies, therefore, may be considered less accurate than the present study or previous detailed studies although the possibility of intraspecific variation in the karyotypes can not be completely disregarded as *Cryptomeria* is a morphologically variable tree (DALLIMORE *et al.*, 1966).

MEHRA and KHOSHOO (1956), MEHRA and ANAND (1979), and SOMEGO and KIKUTI (1980) recognized the presence of a long primary constriction in one chromosome pair. TODA (1977, 1979 a, b, 1980, 1981 a, b) observed this unusual feature but considered it to be a secondary constriction rather than the primary constriction. TODA considered the primary constriction to be located between the long arm and a rounded, short arm referred to as a "Köpfchen", which is the short arm of an acrocentric chromosome (KIYAHARA *et al.*, 1936). This feature can be observed in the second chromosome from the left in Figure 2 b. However, the results of the present study do not agree with TODA's karyological interpretation in this particular instance. The probable position of the kinetochore in the marker chromosomes is indicated by the connected kintonema within the unusual region at the last stage before chromosome separation in meta-anaphase (Fig. 2 a). Inspection of TODA's figures indicates that the "Köpfchen" is a large, inconsistently appearing chromomere in the long arm or could possibly be a feature due to the results of differential reactivity to

staining which has been reported to occur in *Cryptomeria* (SCHLARBAUM and TSUCHIYA, 1981).

MATSUMOTO (1933) reported two chromosome pairs with long kinetochore regions to occasionally occur in cells of *Cryptomeria*. Only one pair was observed in the different clones of the present study or reported in any of the previous studies. However, inspection of the photomicrograph presented by TODA (1981 b) shows two pairs of chromosomes with long kinetochore regions. One pair was the distinctive pair noted in many *Cryptomeria* studies while the other pair, unrecognized by TODA, had a kinetochore region which was comparatively shorter, though the region was definitely longer than a normal primary constriction. It is apparent, therefore, that some populations of *Cryptomeria* have two pairs of chromosomes with long kinetochore regions. More detailed studies are needed to determine if the second marker pair contains kintonemas with chromomeres as in the more distinctive marker chromosomes.

Results of the present study show that the morphology of the long kinetochore regions is variable, although this variation was inconsistent among the marker chromosomes. No observations indicated that the long kinetochore regions of the individual chromosomes were appearing consistently different as in the SAT-chromosomes observed in *Secale cereale* by BOSE (1956, 1957, 1958).

No secondary constrictions were observed in the chromosomes of the different clones in the present study. However, TODA (1979 b, 1981 a, b) has shown indisputable evidence that secondary constrictions are present in some cultivars of *Cryptomeria*. Karyological studies of *Taiwania cryptomerioides* HAY. and *Metasequoia glyptostroboides* HU et CHENG, other members of *Taxodiaceae*, show the presence of marker chromosomes similar to the pair in *Cryptomeria* and an absence of chromosomes with secondary constrictions (SCHLARBAUM and TSUCHIYA, 1984, SCHLARBAUM *et al.*, 1983). Therefore, it is suspected that the long kintono-

chore region, rather than the secondary constriction, contains the nucleolar organization region in addition to the microtubule attachment site. Plant species with unusually long kinetochore regions believed to contain the microtubule attachment site and nucleolar organizer region have been previously reported (HUNZIKER, 1961; KHOSHOO and AHUJA, 1963; KURITA, 1953, 1960; LACOUR, 1950; SATO, 1942). Meiosis of *Cryptomeria* populations with and without secondary constrictions will have to be studied to ascertain the position of the nucleolar organizer region.

Literature Cited

- BOSE, S.: Aberrations in the nucleolar chromosome of inbred rye. *Hereditas* 42: 263–296 (1956). — BOSE, S.: Aberrations in the nucleolar chromosome of inbred rye. Size variation in inbred lines and population plants. *Hereditas* 43: 621–643 (1957). — BOSE, S.: Aberrations in the nucleolar chromosome of inbred rye. Size variation in inbred lines in relation to vigor. *Hereditas* 44: 257–279 (1958). — CHIBA, S.: Triploids and tetraploids of Sugi (*Cryptomeria japonica* D. DON) selected in the forest nursery. *Bull. Govt. For. Expt. Sta. No. 49*: 99–108 (1950). — DALLIMORE, W., JACKSON, A. B. and HARRISON, S. G.: A handbook of *Coniferae* and *Ginkgoaceae*. St. Martin's Press, New York (1966). — ECKENWALDER, J. E.: Re-evaluation of *Cupressaceae* and *Taxodiaceae*: a proposed merger. *Madrono* 23: 237–256 (1976). — FLORIN, R.: The distribution of conifer and taxad genera in time and space. *Acta Horti Berg.* 20: 121–312 (1963). — HAYATA, B.: The *Taxodiaceae* should be divided into several distinct families i.e., the *Limnopytiaceae*, *Cryptomeriaceae*, *Taiwaniaceae*, and the *Cunninghamiaceae*; and further *Tetraclinis* should represent a distinct family, the *Tetraclinaceae*. (Japanese with Latin botanical description). *Bot. Mag.* 46: 24–27 (1932). — HIDA, M.: The systematic position of *Metasequoia*. (Japanese with English summary). *Bot. Mag.* 75: 316–323 (1962). — HUNZIKER, J. H.: Estudios cromosomicos en *Cupressus* y *Libocedrus*. (Spanish with English summary). *Revista de Investigaciones Agricolas* 15: 169–185 (1961). — KANAZAWA, R.: Table of statistics with chromosome of woody plants. (Japanese). *La Kromosomo* 5/6: 249–260 (1949). — KIHARA, H., YAMAMOTO, Y. and HOSONO, S.: Studies on chromosome numbers of plants. (Japanese). 2nd Edition. Yokendo Co. Ltd., Japan (1936). — KHOSHOO, T. N. and AHUJA, M. R.: The chromosomes and relationships of *Welwitschia mirabilis*. *Chromosoma* 14: 522–533 (1963). — KURITA, M.: A study of the chromosomes in *Nothoscordum fragrans* (VENT) KUNTH. *Mem. Ehime Univ.* 1: 55–63 (1953). — KURITA, M.: Nucleolar chromosome with a long centromere. *Rep. Biol. Inst. Ehime Univ.* 9: 1–8 (1960). — KUROKI, Y.: A study related to karyotype of principal conifers, Report of Experimental Plantations. (Japanese). Miyazaki Univ., Fac. Agri. 5: 78–80 (1969). — LACOUR, L. F.: Compound constrictions. *Heredity* 4: 243–246 (1950). — MATSUDA, K. and MIYAJIMA, H.: Basic study related to sterility of *Cryptomeria japonica* D. DON IV, Progress. 87th Meet. Jpn. For. Assoc. 58 (1976). (fide Toda, 1979b). — MATSUMOTO, K.: Über die Chromosomenzahl von *Cryptomeria japonica* D. DON und *Taiwania cryptomerioides* HAYATA. (Japanese with German summary). *Bot. Zool. Tokyo* 1: 1751–1756 (1933). — MATSUMOTO, H.: Chromosome studies on *Trillium kamtschaticum* PALL. XIII. The structure and behavior of the kinetochores. *Cytologia* 11: 369–379 (1941). — MEHRA, P. N. and ANAND, M.: Cytology of calus of *Cryptomeria japonica*. *Physiol. Plant.* 45: 127–131 (1979). — MEHRA, P. N. and KHOSHOO, T. N.: Cytology of conifers. *Indian J. Genet. Plant Breed.* 54: 165–180 (1956). — SAITO, Y. and HASHIZUME, H.: Studies on a triploid specimen of *Cryptomeria japonica* picked out of afforested land. (Japanese with English summary). *Tottori Daigaku Nogyoju Enshurin Hokoku, Bull. Torri. Univ. For. No. 1*: 21–35 (1958). — SATO, D.: Karyotype alteration and phylogeny. V. New type of SAT-chromosomes in *Nothoscordum* and *Nerine*. *Cytologia* 12: 170–178 (1942). — SAX, K. and SAX, H. J.: Chromosome number and morphology in the conifers. *J. Arnold Arbor.* 14: 356–375 (1933). — SAYLOR, L. C.: A karyotypic analysis of selected species of *Pinus*. *Silvae Genet.* 10: 77–84 (1961). — SCHLARBAUM, S. E.: Cytotaxonomic relationships within *Taxodiaceae*. Ph. D. dissertation. Colorado State Univ. (1980). — SCHLARBAUM, S. E. and TSUCHIYA, T.: The chromosome study of giant sequoia, *Sequoiadendron giganteum*. *Silvae Genet.* 24: 23–26 (1975). — SCHLARBAUM, S. E. and TSUCHIYA, T.: Chromosome study of Japanese umbrella pine. *J. Hered.* 67: 65–67 (1976). — SCHLARBAUM, S. E. and TSUCHIYA, T.: Differential reactivity in tree chromosome staining. *J. Hered.* 72: 62–63 (1981). — SCHLARBAUM, S. E. and TSUCHIYA, T.: The chromosome studies of *Cunninghamia konishii* HAY., *Cunninghamia lanceolata* (LAMB.) HOOK., and *Taiwania cryptomerioides* HAY. *Plant Syst. Evol.* 142: 00–00 (1984). (In Press). — SCHLARBAUM, S. E., JOHNSON, L. C. and TSUCHIYA, T.: Chromosome studies of *Metasequoia glyptostroboides* and *Taxodium distichum*. *Bot. Gaz.* 144: 559–565 (1983). — SHIBATA, K., OGOSHI, Y. and NAKATA, G.: Cytological on *Coniferae*. I. Chromosome numbers of some species in *Chamaecyparis*, *Cryptomeria*, and *Pinus*. (Japanese with English summary). *La Kromosomo* 29: 1025–1028 (1959). — SOMEKO, M. and KIKUTI, H.: Cytogenetical studies on artificial triploids and aneuploids of Sugi, *Cryptomeria japonica*. (Japanese with English summary). *Bull. For. Res. Inst.* 310: 171–174 (1980). — STIFF, M. L.: The geographical distribution and cytology of *Coniferales*. Ph. D. thesis. Univ. of Virginia (1952). — TODA, Y.: Karyological studies on triploid *Cryptomeria japonica*. (Japanese with English summary). *La Kromosomo* II-6: 186–190 (1977). — TODA, Y.: Karyotype of *Cryptomeria japonica* D. DON I. (Japanese). *Proc. Trans. Kyushu Branch, Jpn. For. Assoc.* 32: 151–152 (1979a). — TODA, Y.: On the karyotype of *Cryptomeria japonica* D. DON II. *Bull. Fac. Hort., Minamikyushu Univ.* 9: 1–10 (1979b). — TODA, Y.: On the karyotype of *Cryptomeria japonica* D. DON V. *Cryptomeria japonica* D. DON in Kyushu (1). *J. Jpn. For. Soc.* 62: 246–269 (1980). — TODA, Y.: On the karyotype of *Cryptomeria japonica* D. DON VIII. *Cryptomeria japonica* in Kyushu (4). *La Kromosomo* II-24: 707–712 (1981a). — TODA, Y.: On the karyotype of *Cryptomeria japonica* D. DON XIII. *Cryptomeria japonica* D. DON in Kyushu 4. (Japanese with English summary). *La Kromosomo* II-24: 707–712 (1981b). — TSUCHIYA, T.: An improved acetocarmine squash method, with special reference to the modified Rattenbury's method of making a preparation permanent. *Barley Genet. Newslett.* 1: 71–72 (1971). — ZINNAL, I.: Polyploids of a few conifers grown with colchicine. Report of Experimental Plantation, Tokyo Univ. 35: 15–25 (1947) (fide TODA, 1979b). — ZINNAL, I. and S. CHIBA: Naturally occurring tetraploids of *Cryptomeria japonica*. (Japanese with English summary). *J. Breed.* 1: 43–46 (1952).

Clonal selection in *Larix laricina*. I. Effects of age, clone and season on rooting of cuttings

By E. K. MORGENSTERN*, J. M. NICHOLSON* and Y. S. PARK**

(Received 30th September 1983)

Summary

Experiments were conducted with 50 clones of tamarack (*Larix laricina* [Du Roi] K. Koch) of 4 age classes ranging from 3 to 10 years, from which cuttings were taken and

* Faculty of Forestry, University of New Brunswick, Fredericton, N. B. E3B 6C2, Canada.

** Canadian Forestry Service, Maritimes Forest Research Centre, Fredericton, N. B., E3B 5P7.

struck at 4 dates between May and August. The rooting medium was a mixture of peat and vermiculite kept in styro-blocks, and the cuttings were treated with a rooting hormone and irrigated by intermittent mist. Cuttings when struck were characterized by a distinct phenological state. Assessments after 3 months included rooting percent, percent of cuttings with major roots (more than 3 cm long), number of major roots per cutting, length of the longest root, and shoot phenology. Analysis of variance indicated