Karyotype Analysis of Pinus-Group Lariciones')

By L. C. SAYLOR²)

(Received for publication June 27, 1964)

Introduction

Early karyological studies of the genus Pinus describe pine chromosomes as being similar not only within karyotypes but also among the different species. Recent studies have shown, however, that this regularity is not so general as once thought. An example of karyotypic variability is presented in this paper. Karyotypes of 19 species of pine of the Group Lariciones are described. The pines in this Group are found primarily in the Old World where they range from Western Europe to the Far East. Two species are found in the New World. Pinus resinosa is located in northeastern United States and Canada; Pinus tropicalis is restricted to Cuba and the Isle of Pines.

Review of Literature³)

The characteristic haploid karyotype of the genus Pinus was first described clearly by Sax and Sax (1933) as consisting of 12 similar chromosomes. The smallest chromosome is somewhat heterobrachial, and the remaining eleven have approximately median centromeres. Later investigators have wbtained the same general karyotypic pattern in various species. In addition some have also found evidence of interspecific differences in chromosome morphology. Although the magnitude of the evidence in the individual studies generally is not great, an analysis of the collective data discloses certain distinctive features. For example, Asss (1957) examined the karyotypes of 24 trees of Pinus sylvestris and found secondary constrictions in the largest pair of chromosomes. From the photographs and drawings presented in the article, it also appears that the second smallest as well as the smallest chromosome has a submedian centromere. A detailed study of the P. sylvestris karyotype was also made by Natarajan et at. (1961). Their idiogram shows two pairs of heterabrachial chromosomes and secondary constrictions in the largest chromosome pair. In addition, their idiogram shows secondary constrictions in four other pairs of chromosomes.

In a detailed study based on relative arm lengths, SAYLOR (1961) presented evidence of karyotypic variation. The centromeres of the two smallest chromosomes of Pinus resinosa, a member of the Group Lariciones to which P. sylvestris also belongs, were described as being located in a submedian position. Three other species of Lariciones, P. nigra, P. thunbergii, and P. densiflora, also were examined briefly. They possessed the second submedian chromosome characteristic of the karyotype of P. resinosa. These data, combined with the findings of AASS (1957) and NATARAJAN et al. (1961), suggested the submedian location of the centromere of the second smallest chromosome as a diagnostic feature of the karyotypes of species of Lariciones. The present study was designed to investigate this possibility.

Material and Methods

The number of species included in the Group Lariciones varies frwm 12 to 21 depending on the classification system used. The material available for this study included all but two of the 21 species that might be classified as members of this Group. All attempts to obtain material of Pinus hwangshanensis Hsia (found only in China) have failed thus far. Three seed lots of P. merkusii DeVries were acquired, but none of the seeds were viable. Species identification of the material studied was primarily according to information provided by the seed donors. All seedlings used are being grown for positive identification when sufficiently mature.

The karyotypes were determined by studying mitotic chromosomes from squash preparations of root-tip meristems of young seedlings. The root-tips were pretreated in oxyquinoline (0.3 g/l.) for 24–36 hours at 12° C, fixed in 3:1 ethyl alcohol-acetic acid for 1–4 hours, hydrolyzed in 1 N HCl for 10–15 minutes at 60° C, and stained in acetocarmine (Saylor, 1961).

The chromosomes were drawn at 2000× magnification using a projection apparatus. Measurements were made on the drawings. To facilitate studying and comparing the individual karyotypes, the chromosomes were numbered and arranged from 1-12 by descending order of lengths of the a (short) arm. Divergence of the lengths of the b (long) arms from a normal descending sequence and the locations of the centromeres and secondary constrictions were used as diagnostic features of the karyotypes. It was not possible to identify positively each of the chromosomes because the lengths of the a arms of adjacent chromosomes were often very similar. The lengths of the a arms for two and three chromosomes sometimes differed by only 0.5 mm, and the error term for the microtechnical and drawing procedures was of this magnitude. The patterns found in more than 50 per cent of the plates examined for a given species determined the final arrangement of chromosomes. No evidence of intraspecific ~karyotypicvariation has been detected, so a minimum of six drawings from at least four seedlings was considered adequate for defining the karyotypes of the individual species.

Problems encountered in studying secondary constrictions in pine chromosomes have been discussed previously (SAYLOR, 1961). The achromatic regions are often difficult to detect when chromosomes are contracted to the degree that is necessary to study their general morphology. To help overcome this problem, a greater effort was made in this study to select only those plates in which the achromatic regions were evident and the chromosomes were not overly contracted. This proved to be relatively helpful. Secondary constrictions were observed with sufficient repeatability (i.e., in more than 50 per cent of the plates) in the chromosomes of most species to include them as part of the karyotype.

Results and Discussion

The karyotypes of all species examined are basically similar to the general karyotypic pattern presented for the pines by SAX and SAX (1933). The chromosomes within any

¹⁾ Contribution from the Department of Genetics and the Department of Forest Management, North Carolina Agricultural Experiment Station as Journal Paper No. 1766. Supported in part by the National Science Foundation Program in Systematic Biology: Grant G18715.

³) Assistant Professor of Genetics and Forestry, North Carolina State, UNC, Raleigh, North Carolina. The author gratefully acknowledges the Services of his assistant, Mrs. Lynne T. Olsson.

³⁾ A more detailed review of the literature was presented in an earlier paper (SAYLOR, 1961).

Table 1. — Mean arm length in millimeters of chromosomes magnified 2000imes. See note.

Chromo- some	Arms			Arms			Arms			Arms		
	a	b	a/b	a	b	a/b	a	b	a/b	a	b	a/b
		P. densiflor	·a	P	. heldreic	hii		P. insulari	is		P. khasya	****
1 2 3 4 5 6 7 8 9 10 11	12.7 12.0 11.7 11.5 11.2 11.1 10.9 10.6 10.4 9.6 7.7 6.3	13.6* 12.8* 11.8 12.5 11.6 12.3 11.4 12.1 11.4 10.3 10.6 8.8	.934 .938 .992 .920 .966 .902 .956 .876 .912 .932 .726	14.5 14.1 13.7* 13.5 13.1 12.8 12.6 12.1 11.9 10.9 9.4 6.6	16.4 15.1 14.1 15.1 13.6 14.9 13.0 13.9 12.8 12.6 13.1	.884 .934 .972 .894 .963 .859 .969 .870 .930 .865 .717	14.5 14.1* 13.8 13.4 13.1 12.8 12.6 12.2 11.8 10.9 8.4 7.2	16.4 15.1 14.2 14.7 14.0 14.7 13.5 14.7 13.2 11.8 13.6 11.0	.884 .934 .972 .912 .936 .871 .933 .830 .894 .924 .618	14.9 14.4 14.0 13.7 13.4 13.0 12.8 12.4 11.9 10.9 8.8 7.7	16.6 15.2 14.5 15.4 14.0 14.8 13.4 14.6 12.9 11.9 13.5 11.9	.898 .947 .965 .890 .957 .878 .955 .849 .922 .916 .652
	F	. luchuens	is	P.	massonia	ana		P. montan	ıa		P. nigra	
1 2 3 4 5 6 7 8 9 10 11	13.3 13.0 12.6 12.5 x 12.3 12.1 11.8 11.4 11.2 10.4* 8.4 7.1	14.4* 13.7 12.9 13.8 12.7 14.2 12.7 12.3 13.3 11.4 11.8 9.6	.924 .949 .977 .906 .969 .852 .929 .927 .842 .912 .712	13.6* 13.0 12.5 12.3 12.1 11.9 11.6 11.3 11.0 10.1 7.9 6.7	14.3 13.4 12.9 13.3 12.5 12.9 12.2 12.7 12.1 11.4 12.0 10.3	.951 .970 .969 .925 .968 .922 .951 .890 .909 .886 .658	12.9 12.4 12.2 11.9 11.7 11.4* 11.0 10.8 10.3 9.5 7.6 6.7	13.4 13.0 12.5 13.0 12.1 12.6 11.9 11.2 12.4 10.5 10.9 9.7	.963 .954 .976 .915 .967 .905 .924 .964 .831 .905 .697	14.4* 14.0 13.6 13.3 13.0 12.9 12.6 12.1 11.8 10.6 8.6 7.1	15.4 14.6* 14.2 13.6* 13.5* 14.3 ² 13.4 12.7 13.0 12.1 11.9 10.8	.935 .959 .958 .978 .963 .902 .940 .953 .908 .876 .723 .657
	i	P. resinoso	ı	P. sylvestris			P. tabulaeformis			P. taiwanensis		
1 2 3 4 5 6 7 8 9 10 11 12	15.1 14.2 13.7 13.4 13.1 12.9 12.7 12.4 12.1 11.4* 8.7 6.8	15.8* 14.9 14.2 14.9z 13.8 14.8z 13.1 14.2 12.6 12.9 10.9	.956 .953 .965 .899 .949 .872 .969 .873 .917 .905 .674	13.7* 13.0 12.7 12.4 12.1 11.8 11.5 11.2 10.8 9.9* 7.9 6.8	14.7 14.2* 13.1 12.8 13.8 12.3 13.0 11.8 12.4 11.2 11.6 10.2	.931 .915 .969 .969 .877 .959 .885 .949 .871 .884 .681	14.5 14.0* 13.9 13.6 13.3 13.1 12.8 12.5 12.0 11.1 8.9 7.4	15.8* 15.1 14.1 14.8 14.1 13.5 14.6 13.2 13.7 12.1* 12.7	.918 .927 .986 .919 .943 .970 .877 .947 .876 .917	15.1 14.5 14.2 13.9 13.8 13.5 13.3 12.8 12.2 11.3 9.0 7.6	15.7 15.1 15.0* 16.0* 14.4 15.0 13.9 14.9 13.7 13.1 13.4 11.0	.962 .960 .947 .869 .958 .900 .957 .859 .890 .862 .672
	P	. thunberg	ıii	F	. tropical	is	P.	yunnanen	sis	ļ P	. halepensi	s
1 2 3 4 5 6 7 8 9 10 11 12	12.6 12.4 12.2 12.0 11.9 11.6 11.4 10.9 10.7 10.1* 7.6 6.3	13.8 13.1 12.4 13.3 12.4z 13.3 12.4 11.4 12.0 11.3 11.4 9.1	.913 .947 .984 .902 .960 .872 .919 .956 .892 .894 .667	15.5* 15.2 14.8 14.4 14.2 14.0 13.7 13.5 13.0 12.0 9.4 7.1	16.6* 15.8 15.3 16.0 14.7 15.4 14.5 16.0 14.4 13.1 14.3 11.5	.934 .962 .967 .900 .966 .909 .945 .844 .903 .916 .657	14.8* 14.1 13.9 13.5 13.3 13.1 12.9 12.4 12.1 10.7 9.1 7.7	15.9 15.1 14.4 14.1 14.0 13.8 14.5 12.9 12.6 12.8 11.3	.931 .934 .965 .957 .924 .936 .935 .855 .938 .849 .711	16.5 15.5 15.1 14.8 14.5 14.0 13.8 13.6* 13.2 12.4 9.7 7.4	17.3 17.1 16.5 15.7 15.2 16.1 15.5 14.4 16.2 14.5 15.2 13.7	.954 .906 .915 .943 .954 .870 .890 .944 .815 .855 .638
	P. brutia		P. pityusa		P. pinaster							
1 2 3 4 5 6 7 8 9 10 11	16.7 15.8* 15.4 15.1 14.9 14.7 14.4 14.2 13.8 12.9 9.9 8.3	17.7 16.9 16.0 17.3* 15.6 17.0 15.8 14.9 15.8 14.7 15.3 14.3	.944 .935 .962 .873 .955 .865 .911 .933 .873 .878 .647	17.5 16.9 16.3 16 1 15.7 15.5 15.3 14.7 14.1 13.5 10 5 8.5	18.8* 17.8 17.7 16.6 17.2 16.5 16.9 15.6 16.5 15.6 16.3 15.0	.931 .949 .921 .970 .913 .939 .935 .942 .854 .865 .644	12.9* 12.4 12.1 11.7 11.6* 11.4 11.2 11.0 10.6 9.6 8.4 6.6	13.9 13.2* 12.5 12.2 12.9 12.0 11.8 12.5 11.1 10.9 11.1	.928 .939 .968 .959 .950 .949 .880 .955 .881 .754			

Note: -x, *, and z denote secondary constrictions in the distal, medial, and proximal portion of the arms, respectively.

given complement are similar in size and general morphology. They gradually decrease in length, and they possess either median or submedian centromeres. (Chromosomes with median and submedian centromeres are defined by short-arm: long-arm ratios of 0.75-1.00 and 0.50-0.75, respectively.) The smallest chromosome is always heterobrachial.

The mean lengths of the chromosome arms (*Table 1*) do not provide critical information about differences in chromosome lengths among the various species. The apparent specific differences most likely resulted from unequal pretreatment periods, or from differential reaction of the chromosomes to the pretreatment, or both.

The two smallest chromosomes in the haploid complement of every species of Lariciones examined possess submedian centromeres (Tables 1 and 2; Figures 1 and 2). The centromeres of the ten remaining chromosomes are all median. In my overall cytogenetic study of this genus, karyotypes have been determined for 39 species from eight of the twelve Groups listed by Shaw (1914 and 1924). Thus far, only the species of Lariciones have the centromeres of both of the smallest chromosomes located in a submedian position (Tables 2 and 4). This evidence confirmes the earlier findings of Aass (1957), Natarajan et al. (1961), and Saylor (1961) and suggests the existence of inter-Group differences in chromosome morphology.

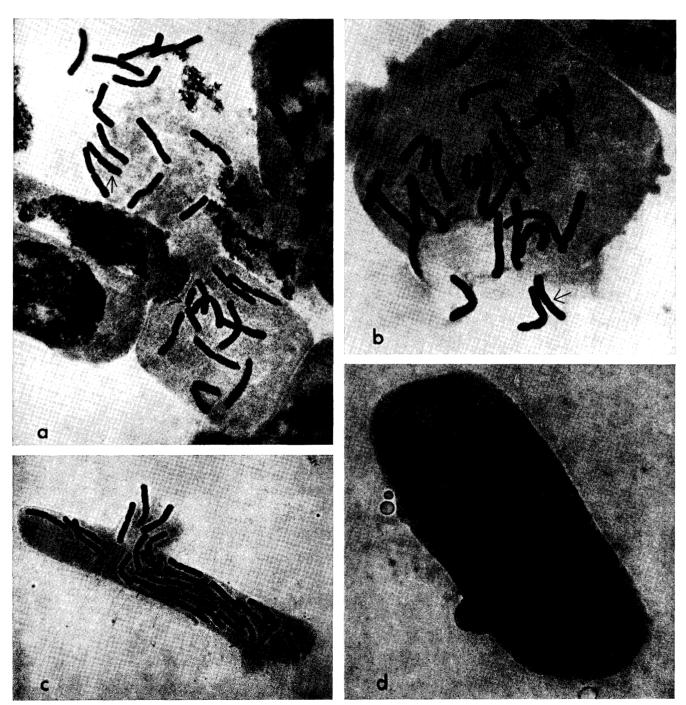


Figure 1. — Chromosomes from aceto-carmine preparations of root-tip meristems. The short and long arrows denote respectively the smallest and second smallest chromosomes, both of which are heterobrachial. (a) P. tropicalis (1025×); (b) P. resinosa (1225×); (c) P. heldreichii (780×); (d) P. sylvestris (1125×).

Table 2. — Diagnostic features of karyotypes of species of Lariciones obtained by arranging the chromosomes according to descending order of the α (shorter) arm lengths as in Table 1.

Species	Plates drawn	Plants	Chromosomes showing ex- ceptions to the descending order in b (longer) arm lengths	Chromo- somes with most median centro- meres	Chromo- some eleven a/b	Chromo- some twelve a/b
P. densiflora Sieb. & Zucc.	10	6	4, 6, 8, 11	3, 5	.726	.716
P. heldreichii Christ.	6	4	4, 6, 8, 11	3, 7	.717	.559
P. insularis Endlich	6	4	4, 6, 8, 11	3, 5	.618	.654
P. khasya Royle	6	4	4, 6, 8, 11	3, 5	.652	.647
P. luchuensis Mayr	8	4	4, 6, 9, 11	3, 5	.712	.740
P. massoniana Lamb.	6	5	4, 6, 8, 11	2, 3	.658	.650
P. montana Miller	7	5	4, 6, 9, 11	3, 5	.697	.691
P. nigra Arnold	6	6	6, 9, 11	4, 5	.723	.657
P. resinosa Air.	13	8	4, 6, 8, 11	3, 7	.674	.624
P. sylvestris L.	13	8	5, 7, 9, 11	3, 4	.681	.667
P. tabulaeformis Carriere	6	4	4, 7, 9, 11	3, 6	.701	.691
P. taiwanensis HAYATA	7	4	4, 6, 8, 11	1, 2	.672	.691
P. thunbergii Parl.	6	6	4, 6, 9, 11	3, 5	.667	.692
P. tropicalis Morelet	8	5	4, 6, 8, 11	3, 5	.657	.617
P. yunnanensis Franchet	7	4	5, 8, 11	3, 4	.711	.681
P. halepensis Miller	6	6	6, 9, 11	1, 5	.638	.540
P. brutia TEN.	6	6	4, 6, 9, 11	3, 8	.647	.580
P. pityusa Steven	10	7	5, 7, 9, 11	2, 4	.644	.567
P. pinaster AIT.	10	6	5, 8, 11	3, 4	.754	.647

Table 3. — Number of species in Table 2 having common patterns of b arm lengths and of location of metacentric chromosomes.

b arm patter	Chromosomes with most median centromere			
Sequence	Species	Positions	Species	
4, 6, 8, 11	8	1, 2	1	
4, 6, 9, 11 Either of above	4	1, 5	1	
with one change	3	2, 3	1	
Other	4	2, 4	1	
		3, 4	3	
		3, 5	7	
		3, 6	1	
		3, 7	2	
		3, 8	1	
		4, 5	1	

Table 4. — Short-arm: long-arm ratio of the second smallest chromosome (number eleven) for species of *Pinus* as arranged by Shaw (1924).

Species	Ratio
Group Strobi	
P. flexilis James	.839
P. strobus L.	.875
P. lambertiana Dougl.	.832
P. flexilis James	.839
Group Balfourianae	
P. aristata Engelm.	.830
Group Longifoliae	
P. canariensis Smith	.769
Group Pineae	
P. pinea L.	.860
Group Australes	
P. palustris Mill.	.944
P. taeda L.	.920
P. caribaea Morelet	.816
P. elliottii Engelm.	.929
P. ponderosa Laws.	.807
P. washoensis Mason and Stockwell	.824
P. jeffreyi Grev. and Balf.	.775
P. glabra Walt.	.864
P. engelmanii CARR.	.843
P. cooperi Blanco	.812
Group Insignes	
P. virginiana Mill.	.878
P. clausa (Chapm.) Vasey	.915
Group Macrocarpae	
P. coulteri D. Don	.803
P. torreyana PARRY	.872

The karyotypes of the species within the Group Lariciones are not readily distinguishable from each other, although subtle interspecific differences possibly exist. Patterns of length of the b arms were determined after arranging the chromosomes of a given karyotype in descending order according to the lengths of the a arms (Table 2). The locations of the two chromosomes with the most median centromeres also were compared. According to these features, different karyotypes exist within this Group, although many of the karyotypes are similar. For example, several species appear to have a common karyotype (Tables 2 and 3; Figure 3) in which the chromosomes in positions 4, 6, 8, and 11 commonly interrupt the descending order of the b arm lengths and in which the most median centromeres occur in positions 3 and 5. Considering the process of speciation these results are not unexpected, and this may be the basic pattern from which the other species in the Group have evolved. If this postulation is true, common patterns may also exist within each of the other Groups.

The location of secondary constrictions is another feature that may be used in determining differences among karyotypes. In 18 of the 19 species examined, secondary constrictions were observed with sufficient repeatability to be included as part of the karyotypes (Table 1). The number of chromosomes in a karyotype having secondary constrictions ranged from one to five. The most consistent results were observed for *P. massoniana*; secondary constrictions were found in the same region of chromosome number 1 in every plate drawn of this species. A similar situation was found for chromosome number 1 in eight out of ten plates of *P. pinaster*.

The location of the achromatic regions was sufficiently variable to suggest that this evidence is not absolutely conclusive. The results are considered accurate enough for indicating where secondary constrictions are most probably located. The value of secondary constrictions as a diagnostic feature of pine karyotypes remains questionable, because the numbers and positions of these regions reported by different investigators frequently vary for any one species. For example, Aass (1957) observed an achromatic region in the longer arm of the largest chromosome of the *P. sylvestris* complement. However, Natarajan et al.



Figure 2. — Projection drawings of chromosomes obtained from aceto-carmine squash preparations of root-tip meristems. The short and long arrows denote respectively the smallest and second smallest chromosomes, both of which are heterobrachial. (a) P. halepensis (Reduced to 1200×); (b) P. pinaster (Reduced to 1200×).

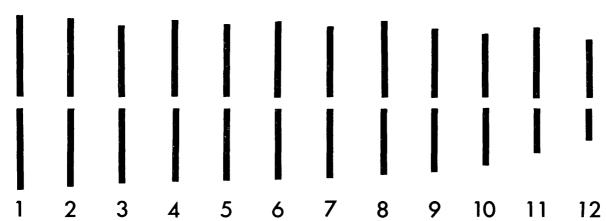


Figure 3. — Idiogram representing the basic chromosome pattern characteristic of species of Lariciones. Chromosomes arranged in descending order of length of the shorter arm. (See text for further explanation.)

(1961) placed it in the short arm of the same chromosome, and they described four other secondary constrictions that Aass did not report. The results of this study support the findings of Natarajan et al. for chromosomes 1 and 10, but differ for chromosomes 2, 6 and 7. An achromatic region was found in the longer arm rather than the shorter arm of chromosome 2. No secondary constrictions were observed in chromosomes 6 and 7.

Species of pine have been classified according to several different criteria such as seed and cone characteristics (Shaw, 1914), needle numbers (Pilger, 1926), and crossability patterns (Duffield, 1952). In general, there is little disagreement among the different classifications with respect to the *Lariciones*. The present study provides karyological data substantiating the interrelationship of the species in this Group. In addition it provides information that should help clarify the taxonomic position of four controversial species.

Pinus halepensis has been placed in various Groups, depending on the characteristics used in the classification. Shaw placed this species in the Group Insignes, while Pilger assigned it to the Section Banksia. Duffield concluded, however, from morphological, anatomical, and biochemical data that P. halepensis is more closely related to species of Lariciones. The presence of the heterobrachial eleventh chromosome, characteristic of the Lariciones, in every examined plate of P. halepensis supports Duffield's conclusion. Similar evidence was found for P. brutia and P. pityusa, two taxa often considered varieties of P. halepensis, which further corroborates the relationship of P. halepensis to the other Lariciones.

Pinus pinaster is another species whose taxonomic position is questionable. Shaw considered this pine to be more closely related to species of *Insignes*, while Pilger and

⁴⁾ PILGER'S Sections are taxonomically equivalent to the Groups of Shaw and Duffield.

Duffield placed it in the *Lariciones*. The karyological data for this species are not so easily interpreted as are the others, because the average value of the short-arm:long-arm ratios for the eleventh chromosome is slightly greater (0.754) than the upper limit of the submedian category; four of the ratios are just below 0.750 and six are slightly above. However, karyotypes for 20 species from seven other Groups have been determined, and only two of these species have a ratio below 0.800 for the eleventh chromosome (*Table 4*). The value for *P. pinaster* is so close to the arbitrary limit of the submedian category that it resembles the species of *Lariciones* more than those in any of the other Groups studied, including *Insignes*.

Summary

Karyotypes are described for 19 species of pine of the Group *Lariciones*. All species had a similar but distinct karyotype characteristic of the Group. The haploid *Lariciones* karyotype differs from those of other Groups by containing two heterobrachial chromosomes instead of one.

Karyological evidence supports the inclusion of *Pinus halepensis*, *P. brutia*, *P. pityusa*, and *P. pinaster* in the Group *Lariciones*. The karyotypes of these species are very similar to those of the generally accepted species of *Lariciones*, because they possess the heterobrachial eleventh chromosome characteristic of this group.

Résumé

Titre de l'article: Analyse des karyotypes des pins du groupe Lariciones.

L'auteur décrit les karyotypes de 19 espèces de pin du groupe Lariciones. Toutes les espèces présentent le karyotype caractéristique du groupe; chaque karyotype est cependant distinct. Le karyotype haploïde des Lariciones diffère de ceux des autres groupes: il comporte deux chromosomes hétérobrachiaux au lieu d'un.

L'analyse karyologique permet d'inclure dans le groupe Lariciones les espèces suivantes: Pinus halepensis, P. brutia, P. pityusa et P. pinaster. Ces espèces sont très semblables à ceux des espèces généralement considérées comme Lariciones, parce qu'elles ont le onzième chromosome hétérobrachial.

Zusammenfassung

Titel der Arbeit: Karyotypen-Analyse der Kiefern-Gruppe der Lariciones.

Für 19 Kiefernarten der Gruppe der Lariciones sind die Karyotypen beschrieben worden. Sie waren alle ähnlich, aber characteristisch für diese Gruppe. Der haploide Lariciones-Karyotyp unterscheidet sich durch das Vorkommen von 2 heterobrachialen Chromosomen, anstatt einem solchen bei anderen Kiefern-Gruppen.

Der karyologische Befund verlangt die Zuordnung von Pinus halepensis, P. brutia, P. pityusa und P. pinaster zur Gruppe der Lariciones. Die Karyotypen dieser Arten sind denen der bisherigen Lariciones sehr änhlich, weil sie das für diese Gruppe charakteristische heterobrachiale 11. Chromosom besitzen.

List of References

AASS, I.: A cytological analysis of Scots pine (Pinus sylvestris L.) from Skjåk, Norway. Meddel. Norske Skogfosøksvesen 14: 96—109 (1957). — Duffield, J. W.: Relationships and species hybridization in the genus Pinus. Z. Forstgenetik Forstpfl.züchtung 1: 93—97 (1952). — Natarajan, A. T., Ohba, K., and Simak, M.: Karyotype analysis of Pinus sylvestris. Hereditas 47: 379—382 (1961). — Pilger, R.: Gymnospermae. In: Die natürlichen Pflanzenfamilien. Von Engler und Prantl. 2. Aufl., 13. Band. Leipzig (1926). — Sax, K., and Sax, H. J.: Chromosome number and morphology in the conifers. Jour. Arnold Arboretum 14: 356—375 (1933). — Saylor, L. C.: A karyotypic analysis of selected species of Pinus. Silvag Genetica 10: 77—84 (1961). — Shaw, G. R.: The genus Pinus. Arnold Arboretum Publ. 5, 1914. — Shaw, G. R.: Notes on the genus Pinus. Jour. Arnold Arboretum 5: 225—227 (1924).

Effects of Inbreeding in Red Pine, Pinus resinosa Ait.1)

By D. P. FOWLER²)

(Received for publication May 1, 1964)

Introduction

"It would seem unwise to conclude that red pine has existed as a specific entity since the Cretaceous, for its Tertiary record is unknown, but the morphological evidence is temptingly suggestive." (Pierce, 1957.)

Deposits assigned to the early Upper Cretaceous Dakota series have yielded cones and needles (Chaney, 1954) and pollen (Pierce, 1957) which are almost identical to those of contemporary Pinus resinosa. If these Cretaceous species (Pinus clementsii Chaney and/or Pinus resinosipites Pierce) are in fact red pine or close relatives of red pine, then the ancestry of this species extends back at least a hundred million years.

Red pine is the only North American species of the Lariciones group. Pinus tropicalis Mor., which occupies a

limited range in Cuba and on the Isle of Pines, is the only other American representative of this group which is largely confined to Europe and Asia (Shaw, 1914).

Attempts to hybridize red pine with other species in the Lariciones group have, until recently, been unsuccessful. Duffield and Snyder (1958) reported a successful cross of Pinus nigra Arn. \times Pinus resinosa, but thus far, attempts to repeat this cross have been unsuccessful.

The natural range of red pine is presented in *Figure I* and is based primarily on the map of Rudolf (1957), with a few changes for Ontario and Manitoba. The range map is somewhat deceiving in that it indicates a relatively continuous distribution. Along the range limits, the distribution of red pine is actually disjunct and, in some cases, distances of 100 miles or more may separate individual locations. Outlying populations are found in West Virginia and northern Illinois (Rudolf, 1957) as well as in northern Ontario (Haddow, 1948), and Manitoba (Vaartaja, 1962).

Cool to warm summers, cold winters, moderate rainfall, sandy soils and undulating topography are characteristic of the red pine habitat. This species occurs in areas of acid

 $^{^{\}rm i}$) Contribution 64 - 5. Ontario Department of Lands and Forests. This paper is the first of a series of papers based on a dissertation submitted to the Graduate School of Yale University as partial fulfillment of the requirements of the PhD degree in 1963.

⁵⁾ Research Scientist, Research Branch, Ontario Department of Lands and Forests, Maple, Ontario, Canada.